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REVIEW



Mast cell activation: beyond histamine and tryptase

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ABSTRACT

Introduction: Mast cells are found in all tissues and express numerous surface receptors allowing them to sense and respond to allergic, autoimmune, environmental, neurohormonal, pathogenic and stress triggers. Stimulated mast cells are typically called 'activated' but the mechanisms involved and the mediators released can vary considerably. Mast cell activation diseases (MCADs) include primary, secondary and idiopathic conditions, especially mast cell activation syndrome (MCAS), but mast cells are activated in many other disorders making the diagnosis and treatment challenging.

Areas covered: Mast cells can release numerous biologically active mediators, some of which are prestored in secretory granules while others are newly synthesized and released without degranulation. Most of the emphasis has so far been on secretion of histamine and tryptase, which do not explain all the multisystemic symptoms experienced by patients with MCADs. As a result, drug development has focused on antiproliferative therapy or blocking the action of individual mediators and not on inhibitors of mast cell activation.

Expert opinion: Activated mast cells are involved in the pathogenesis of MCADs, but also in other disorders making appropriate diagnosis and treatment challenging. The definition of mast cell activation should be expanded beyond histamine and tryptase, with an emphasis on better detection and treatments.

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1. Introduction

Mast cells [1–5] have existed for almost 500 million years [6,7] in many different species, including invertebrates and lower vertebrates (e.g. frogs, lizards, zebrafish) [8–10], suggesting that they served a critical role acting as multifunctional effector cells [11] or pluripotent 'immunoendocrine master players' [12].

Previous reviews have discussed the role of mast cells in health and disease not only in allergies [1,13–21], but also in immunity and inflammation [19,22–24]. In mammals, mast cells derive from hematopoietic CD34+ precursors [25–27], travel in the circulation as precursor cells, and express the surface antigens CD117 (KIT), a tyrosine kinase receptor which is the ligand for stem cell factor (SCF) [28]. In systemic mastocytosis, mast cell precursors also express CD3 and CD25. Mast cells are typically activated by allergens crosslinking specific immunoglobulin E (IgE) bound to its high-affinity surface Fc epsilon receptor 1 (FcεRI) [29,30]. IgE probably developed some 300 million years ago, and interesting that wild animals have much higher circulating IgE than humans, probably as a defense against parasites [31]. Interestingly, circulating basophils also express FcεRI and may have co-evolved with mast cells, but their pathophysiology is quite distinct [32,33]. Even though mature mast cells reside in the tissues, they probe the blood vessel lumen by extending filopodia through endothelial gaps, capturing circulating Ig [34,35]. Contrary to early research, fetal mast cells can bind maternal circulating IgE and contribute to postnatal allergic responses [36]. Surprisingly, prenatal stressful events increased umbilical

cord blood IgE [35]. Mast cells can be stimulated by a plethora of other triggers indicating mast cell activation beyond IgE [37], and also the role of IgE beyond mast cells [38].

Mast cells are located in all tissues [39] perivascularly [26,40,41]. The chemokines C-C motif chemokine ligand 2 (CCL2) and CCL5/Regulated on Activation, Normal T Expressed and Secreted (RANTES) are potent chemoattractants for mast cells [42,43]. Mast cells 'mature' under the influence of local micro-environmental factors [44,45] such as interleukin-4 (IL-4), IL-6 and nerve growth factor (NGF) [46]. In fact, mast cells synthesize and release NGF, themselves [47]. Mast cells can assume different phenotypes [48], and have typically been divided based on their secretory granule content of serine proteases [49–51]. Mucosal mast cells (MMC) contain only chymase and connective tissue mast cells (CTMC) contain both chymase and tryptase [52]. However, it has never been shown conclusively that both these proteases are stored in the same secretory granules, and evidence had also indicated there may be more than one form of chymase [53]. Mast cell granules also store tissue remodeling enzymes, such as carboxypeptidase A3 (CPA3) and matrix metalloproteinases (MMPs) [54–57].

Mast cell subtypes can also be further distinguished based on the granule content of biogenic amines [58] and mucopolysaccharides such as chondroitin sulfate and heparin [1]. However, mast cell phenotyping may be modified the discovery of additional unique mast cell molecules [59,60] and expression of various surface receptors [61,62]. Figure 1 shows different models of release of mediators from mast cells that should be considered under the term 'activation.'

Article highlights

- Mast cells have many different phenotypes that can change given the specific tissue microenvironment
- Mast cells synthesize, store and release hundreds of mediators in response to allergic, environmental, neuroimmune, stress and toxic triggers
- Release of histamine and tryptase are limited to degranulation, while other angiogenic, fibrotic, anti-inflammatory, proinflammatory and neurotoxic molecules can be released without degranulation
- Mast cell 'activation' should not be limited primarily to histamine and tryptase release
- Mast cell activation syndrome still requires serum tryptase elevation associated with an episode, but many more patients may qualify for mast cell activation, unspecified or for other mast cell activation disorder
- Activated mast cells are involved in many other conditions making diagnosis challenging and treatment difficult
- Treatment should focus on inhibition of mast cell activation, in addition to inhibiting the effects of selective mast cell mediators.

2. Mast cell activation disorders (MCADs)

Understandably, many reviews of mast cell activation diseases (MCAD)s focused on mastocytosis [39,63–67]. Briefly, MCADs can be divided as follows 1. Primary [e.g. mastocytosis, monoclonal mast cell activation disorder (MMAS)]. 2. Secondary [e.g. allergies, asthma, cancer, urticarias], and 3. Idiopathic [e.g. anaphylaxis, chronic spontaneous urticaria, mast cell activation syndrome (MCAS), mast cell activation, unspecified]. Sometimes, Primary MCADs are referred to as 'intrinsic' (1), while the Secondary MCADs as 'extrinsic.'

MCADs have different ICM1-CM diagnostic codes (Table 1) described in more detail by Valent et al. [68].

MCADs can be confusing [69–74] because there are over 30 other conditions in which activation of mast cells is suspected of being involved such as irritable bowel syndrome (IBS) [75] or autism spectrum disorder (ASD) [76] (Table 2). Moreover,

there are also other conditions that mimic MCADs, but probably do not involve mast cells (Table 3). Moreover, there are conditions that are frequently comorbid with MCADs such as postural tachycardia syndrome (POTS) and hypermobile Ehlers-Danlos syndrome (EDS) [77–79] that together with MCAS have been called the 'triad' [80]. It was recently reported that dysautonomia was documented in most patients with MCAS [81]. Finally, many patients with MCADs are more sensitive to excipients such as additives, colorings and preservatives found in many foods, cosmetics, drugs and supplements [62].

The diagnosis of atopic diseases in general, and MCADs, in particular, depends on clinical history, the response to medications that can reduce mast cell-related symptoms, as well as the elevation of blood and urine biomarkers (Table 4) [82,83]. However, there are instances where mast cell involvement remains unrecognized as in patients with alpha-gal syndrome, which involves specific IgE to the oligosaccharide galactose- α -1,3-galactose (alpha-gal), and is often associated with tick bites, but is also found in mammal-derived ingredients commonly found in numerous cosmetics, drugs and supplements [84].

The definition of MCAS has generated considerable controversy [85–88]. There have been numerous papers presenting consensus criteria for MCAS, the most recent of which was by Dr. Gulen et al. that stressed the importance of incidence-associated elevation of serum tryptase levels and downplayed the significance of urine histamine and prostaglandin D2 (PGD2) metabolites [89]. These criteria translate to many fewer cases of idiopathic MCAS than suspected [90]. These criteria were subsequently slightly 'adjusted' to describe "a subset of patients with no underlying disease, no known trigger of mast cell activation and no specific criteria as of yet [68]. The original description of MCAS by Dr. Akin et al. [91] included elevated 24-hour urine 11β -PGF $_2\alpha$, which was more

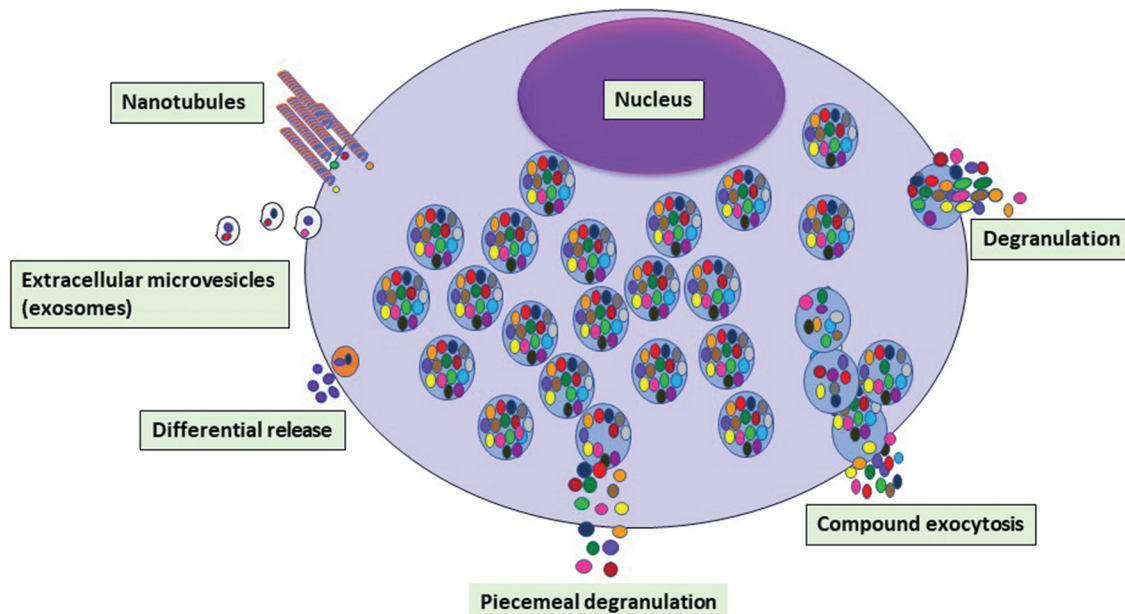


Figure 1. Schematic representation of different models of release of mediators from mast cells that should be considered under the term 'activation'.

Table 1. ICD10-CM diagnostic codes for MCADs (excluding mast cell neoplasms and mast cell leukemia).

C96.21	Aggressive systemic mastocytosis
D47.02	Systemic mastocytosis
D47.01	Cutaneous mastocytosis
D89.4	Mast cell activation syndrome and related disorders
• 40	Mast cell activation, unspecified
• 41	Monoclonal mast cell activation syndrome
• 42	Idiopathic mast cell activation syndrome
• 43	Secondary mast cell activation
• 49	Other mast cell activation disorder

Table 2. Conditions suspected of involving activation of mast cells.

- Alpha-Hypertyptasemia
 - Amyotrophic lateral sclerosis (ALS)
 - Alzheimer's disease (AD)
 - Attention deficit/hyperactivity disorder (ADHD)
 - Autism spectrum disorder (ASD)
 - Chronic inflammatory response syndrome (CIRS)
 - Coronary hypersensitivity (Kounis) syndrome
 - Dysautonomia syndrome
 - Endometriosis
 - Eosinophilic esophagitis/gastroenteritis
 - Familial hyper IgE syndrome
 - Interstitial cystitis/Painful bladder syndrome (IC/PBS)
 - Irritable bowel syndrome (IBS)
 - Long-COVID syndrome
 - Macrophage activation syndrome
 - Migraine headaches
 - Multiple chemical sensitivity syndrome
 - Multiple sclerosis
 - Multisystem inflammatory syndrome (MIS)
 - Myalgic encephalomyelitis/Chronic fatigue syndrome (ME/CFS)
 - Neurofibromatosis
 - Pediatric acute neuropsychiatric syndrome (PAN)
 - Periodontal disease
 - Post-Lyme syndrome (PLS)
 - Psoriasis
 - Sick building syndrome (SBS)
 - Toxic mold syndrome (TMS)
 - Traumatic brain injury (TBI)
 - Vitiligo
-

Table 3. Diseases confused with mast cells disorders.

- Angioneurotic edema
 - Carcinoid syndrome
 - Cryopyrin diseases
 - Dysautonomia
 - Scleroderma
 - Scombroid (histamine) toxicity
 - Vasovagal attack
-

Table 4. Laboratory tests for diagnosis of atopic conditions.

Blood
• IgE, IgG ₁ , IgG ₄
• Immune IgE (RAST for alpha-gal, casein, gluten, dust mites, fungi, grass, pollen)
• Anti-IgE receptor antibody (basophil activation test)
• CCL2, CXCL8 (IL-8)
• Food Intolerance Panel (immune IgG ₄)
• IL-4, IL-6, IL-31
• Tryptase
Urine 24 hours or first-morning void (must be kept and sent cold)
• LTE ₄
• N-Methylhistamine (NMH) or methylimidazole acetic acid (MIA)
• PGD ₂
• 2,3-Dinor-11b-PGF _{2a}

Table 5. Proposed characteristics for a subgroup of MCAS resistant to treatment.

- Presence of neurologic and cognitive symptoms
 - Extreme sensitivity to odors and stress
 - Variable elevated serum levels of:
 - Chymase
 - CRH
 - Eosinophilic cationic protein (ECP)
 - Heparin
 - IL-6
 - IL-31
 - IL-33
 - Matrix Metalloproteinase-9 (MMP-9)
 - MRGPRX2
 - Neurotrophins
 - Osteopontin
 - PAF
 - SP
-

frequently elevated than N-methyl histamine or serum tryptase [92,93], but also the presence of neuropsychiatric symptoms. It was recently reported that patients with hereditary alpha-hyperpotasemia, unlike those with indolent systemic mastocytosis, did not have urine elevations of either N-methyl histamine or 2,3-dinor-11 β -PGF₂ α [94]. A recent review tried to associate some 'non-tryptase urinary and hematologic biomarkers' with MCAS symptoms [95].

The critical issue is not how to define certain syndromes, but how to define mast cell 'activation.' For instance, many patients with MCADs (e.g. allergies, asthma, cancer, urticarias) do not have elevated serum levels of tryptase [96]. This should not be surprising given the fact that mast cells can be stimulated by many neuroimmune- endocrine triggers [97], leading to the release of mediators other than histamine or tryptase [96].

Moreover, a subgroup of MCAS patients have extreme sensitivity to odors and stress, present with neurologic and cognitive dysfunction, and are resistant to treatment with antihistamines and/or leukotriene antagonists (Table 5). Such patients should be

investigated for additional mast cell-associated biomarkers such as IL-6, Osteopontin and vascular endothelial growth factor (VEGF) (Table 4) [98]. A somewhat analogous situation is that of Chronic Spontaneous Urticaria (CSU), in which about 40% of patients are resistant to treatment with antihistamines, but was characterized by elevated blood levels of the mast cell mediator platelet-activating factor (PAF) [99], which is also involved in inflammation [100]. In fact, elevated levels of PAF had been reported to better reflect the severity of anaphylaxis than either histamine or tryptase [101].

Many MCAS patients describe sudden mast cell reactivity to multiple triggers (old and new) following a major stressful event such as a death in the family, major surgery or extensive trauma. This hinged reactivity may be mediated via the action of corticotropin-releasing hormone (CRH) secreted under stress that has been repeatedly shown to worsen atopic conditions [102] and MCDAs. The main receptor for CRH [receptor-1 (CRHR-1)] was shown to be expressed in mast cells of two patients with mastocytosis [103], and functional on human mast cells leading to the selective release of VEGF without tryptase [104]. Hence, addressing stress should be considered in the treatment of MCADs.

3. Pathophysiology of mast cells

In addition to FcεRI, mast cells express multiple surface receptors for a variety of stimuli (Table 6) [97,105], including receptors for sex hormones [106]. Mast cells also can synthesize hormones [107] and neuropeptides such as CRH [108], as well as the peptides hemokinin-1 (HK-1) [109], NGF [110], neurotensin (NT), substance P (SP) [111] and other neurotrophins [112]. Mast cells in the pineal and the hypothalamus may also be involved in circadian rhythms [113–115]. There are also a number of putative inhibitory receptors (Table 7) [116], such as CD300a [117–120], as well as Siglec-6 [121], Siglec-7 [122,123] and Siglec-8 for which an anti-Siglec-8-antibody was recently reported in a Phase II study to improve antihistamine-resistant CSU [124].

The mode and extent of mast cell responsiveness ultimately depend on the interplay between stimulatory and inhibitory signaling pathways (Tables 6 and 7) [125].

Mast cells are also triggered by non-IgE stimuli [19,126,127] and by additional ligands [128], including complement fragments [129,130] neuropeptides [97], such as CRH [131], HK-1 [109], NGF [110], NT [132], SP [133] and somatostatin [134,135] via high-affinity receptors, as well as by many cationic compounds through the low-affinity G-coupled receptor MRGPRX2 [136]. This latter process is distinct from that utilizing the FcεRI and may lead to the release of different mediators. Responses due to degranulation of mast cells via activation of MRGPRX2 and FcεRI were reported to be additive [137]. Moreover, the alarmin IL-33 was shown to amplify production of pro-inflammatory cytokines via activation of either MRGPRX2 or FcεRI [138]. Mast cells can also be stimulated via activation of toll-like receptors (TLRs) [139,140] pathogens, toxins [141], and pathogens [142–144] as well as viruses [145,146] including severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) [147–153]. Mast cells also communicate with T-cells during immune response [154,155]. In fact, mast cells can function as antigen-presenting cells [156] and induce the maturation of dendritic cells [157].

Table 6. Main mast cell stimulatory receptors.

Receptor	Ligand/Stimulus
A2A, A2B, A3	Adenosine
Ach-m	Acetylcholine
ACTH-R	Adrenocorticotrophin
ACE2	Angiotensin 2
Beta2-Adrenoreceptor	Adrenaline
Cannabinoid CB2 receptor	2-arachidonoyl-glycerol, anandamide
C3a, C5a	Complement
CD3, CD25,CD34,CD66,CD326	Unknown
CXCR1–4	Chemokines
CD47	Integrins
CD48	2B4
CD117 (KIT)	Stem cell factor
CD226/DNAM-1	Nectin-2 (CD112)
CD300	Eosinophilic Cationic proteins
CLR (calcitonin receptor-like receptor)	Calcitonin gene related peptide)
CRHR-1, 2	CRH, urocortin
EMR2	Vibrations
Estrogen receptors A,B	Estrogens
ETA, B	Endothelin-1
FcαRI (CD89)	IgA
FcεRI	IgE
FcγRI, RIIA, RIIIB, RIIIC	IgG
GABA-A, B, C	Benzodiazepines, gamma-aminobutyric
NMDAR, AMPAR, and kainate receptors	Glutamate
Heparan sulfate	Bacterial, viral antigens
H1, H2, H3, H4	Histamine
IL-1R1	IL-1β
IL-4 R	IL-4, IL-13
IL-6 R+IL6ST/GP130/IL6-β	IL-6
IL-10R1,2	IL-10
IL-17 R	IL-17
IL-18Rα+IL-18Rβ	IL-18
IL-36 R	IL-38
LDL, VLDL	Apolipoprotein E
Mel1a, Mel1b, MT1, MT2	Melatonin
NGFR (CD271 or p75 neurotrophinR)	Nerve growth, brain-derived neurotrophic factor
MHCI, II	Antigenic peptides and drugs
MRGPRX2	Cationic peptides
NK-1	Substance P, hemokinin-1
NT3	Neurotensin
Opioid receptors	Endorphins, enkephalins
PAF-R	Platelet-activating factor
Protease-activated receptor-2 (PAR-2)	Proteases
Progesterone receptor	Progestins
PTH1R (PTH/PTHrP type 1 receptor)	Parathyroid hormone
Purinoreceptors	ATP
STTR1-STTR5	Somatostatin
ST2	IL-33
TGFBR1,2 and 3	TGFβ
TLR(1–9)	DAMPs, Pathogens
VDR	Vitamin D

Upon stimulation, mast cells rapidly secrete via degranulation multiple mediators [158] that include preformed, granule-stored mediators such as heparin, histamine, tryptase and tumor necrosis factor (TNF) [3]. Histamine has been the main mediator associated with mast cells [159] but is also released from basophils [160]. Interestingly, mast cells can also generate a histamine-releasing peptide from albumin [161], meaning that once stimulated mast cells can release enzymes that can act on albumin and produce a peptide that can further stimulate mast cells. The serine proteases chymase and tryptase are considered to be stored in the same secretory granules. Tryptase has been studied extensively because it is considered an exclusive mast cell-

Table 7. Main mast cell inhibitory receptors.

Receptor	Ligand
Allergin-1	Inhibitory
ALX/FPR2	LXA ₄ (proresolving)
CD200R	CD200 glycoprotein
Glyceraldehyde-3-phosphate dehydrogenase (GAPDH)	Lactoferrin
IL-10RI and IL-10RII	IL-10
IL-18Ra and IL-1R8	IL-37
IL-36 R a	IL-38
Nucleotide converting ectoenzymes (E-NDP1,3,7)	ATP
Peroxisome proliferator-activated receptor gamma (PPAR-γ)	15d-prostaglandin J2 (15d PGJ ₂)
Protein tyrosine phosphatase (PTPs)	Chondroitin sulfate
IRp60	Unknown
Siglec-6,7,8	Unknown
Thrombomodulin	Thrombomodulin (soluble)
TMEM184A	Heparin

associated mediator and is released during anaphylaxis and systemic mastocytosis [162,163], as well as in many cases of MCAS [164]. Tryptase activates the pro-inflammatory protease-activated receptors (PARs) and generate anaphylatoxins (C3a, C5a) [165]. However, there are a number of unexplained findings related to tryptase [166]: (a) Elevated PAF levels are better correlated with anaphylaxis than tryptase or histamine [101]; (b) lack of tryptase elevations in many patients with MCAS [96]; (c) acute increases in serum tryptase were not associated with symptoms in diffuse cutaneous mastocytosis [167]; and (e) mast cell activation in patients with coronavirus disease 2019 (COVID-19) was associated with serum greater elevations of chymase, than tryptase or other mast cell mediators [151]. This latter finding may be possibly due to the preferential activation of pulmonary mucosal mast cells that contain chymase but not tryptase.

Mast cells also secrete newly synthesized mediators 6–24 hours after stimulation (late- [151] phase reaction); these include prostaglandin D₂ (PGD₂) [168], cytokines (IL-5, IL-6, IL-31, IL-33 and TNF-α) and chemokines (CCL2, CCL5 and CXCL8) [4,5,169], as well as PAF [170,171] which has been implicated in micro thromboses [172,173] and inflammation [172]. PAF is the most potent trigger of platelet aggregation known, but it is extremely short-lived making its routine measurement difficult [174]. PAF is involved in allergies [175] such as allergic rhinitis [176–178] immediate and late cutaneous reactions [179], as well as chronic spontaneous urticaria (CSU) [99]. Combination treatment blocking both PAF and histamine by the histamine-1 receptor antagonist rupatadine markedly reduced the severity of peanut-induced anaphylaxis [180]. PAF also stimulates mast cells [170] and eosinophils [181,182], as well as induces IL-6 production [183–185], which in turn stimulates the production of PAF [186,187]. Mast cells also secrete mitochondrial DNA that induces autocrine and paracrine inflammatory responses [188].

4. Selective release of mediators

Mast cells can release specific mediators via differential release mechanisms [189]. It was first shown that was possible for serotonin [190–192], and VEGF [49] without degranulation, but rather via intragranular changes associated with the release of

mediators without the release of histamine or tryptase [193]. It had been also reported that mast cells can release the content of individual secretory granules [194] or individual mediators without degranulation [190]. This process was distinct from ‘piece-meal degranulation’ that had already been reported [195], granule-associated vesicle transport [196], or the release of extracellular vesicles [197–203].

Mast cells could also form antibody-dependent ‘synapses’ for dedicated secretion [203,204] and also communicate via the formation of nanotubes [205]. The ‘alarmin’ IL-33 [96,206,207] stimulates mast cells via activation of their own specific surface receptor, ST2/IL-33, significantly increasing the ability of SP to stimulate the release of VEGF [133,208], IL-31 [209], TNF [210] and IL-1β [211] as well as CCL2 and CXCL8 [212] and other newly synthesized mediators [96]. It was recently reported that VEGF and other angiogenic factors were elevated in the serum of patients with mastocytosis and the degree of elevation correlated with disease severity [213,214]. VEGF is also vasodilatory and was shown to be increased in lesional skin in CSU [213].

IL-33 also augments the release of IL-31 from human mast cells stimulated either by SP or IgE/anti-IgE [112]. Mast cells can release IL-33, themselves [116]. Mast cell-derived IL-1β or histamine-induced release of IL-1β from macrophages [215] can then stimulate mast cells to release IL-6 selectively without degranulation [191,192]. IL-6 is elevated in MCADs and correlated with disease severity [216–218] and is also elevated in COVID-19 [122;123]. In fact, IL-6 promotes an increase in mast cell numbers [219], and is constitutively released in the presence of the D816V-KIT mutation [220]. The mode and extent of mast cell responsiveness ultimately depend on the interplay between stimulatory and inhibitory signaling pathways (Tables 6 and 7).

Secretion of mediators can occur utilizing different signaling [20,221–223] and secretory [221,223] pathways (Figure 1).

5. Epigenetic regulation of mast cell functions and mast cell-associated disorders

Epigenetics refers influence of environmental and behavioral events that alter genes without changing the DNA sequence via DNA methylation, posttranscriptional modifications through non-coding RNAs, histone modifications and nucleosome positioning that affects DNA and chromatin structure [224].

Epigenetic modifications can be suppressed, cured and reversible including in MCAD [225]. Mast cells could undergo epigenetic programming that maintains a stable identity but can undergo significant plasticity to adjust to different environmental challenges [226]. Though the implication of epigenetic changes in the pathogenesis of systemic mastocytosis is not yet clearly known [224], several reports indicated epigenetic modifications may be involved in MCADs [225,227,228]. In fact, epigenetic mechanisms could contribute to the familial type of systemic MCAD [228]. In particular, some mast cell mediators could have autocrine effects. For example, tryptase could catalyze histone clipping as a novel epigenetic mechanism in mast cells [229], and tryptase could regulate the epigenetic modification of histones in mast cell leukemia cells [230].

DNA methyltransferase 3A (DNMT3A) and DNA methylation play a significant role in acute and chronic mast cell activation conditions [231]. Mastocytosis could be treated with inhibitors of histone methyl transferase that can cause cell death by apoptosis [232]. Epigenetic changes in macrocytosis may include mutations in the genes such as TET1, DNA methyltransferase enzyme DNMT3A, ASXL1 and gene methylation in neoplastic cells [224]. Ten-eleven translocation-2 (TET2) is an epigenetic regulator and its expression is induced in response to both acute and chronic activation of mast cells [226,233]. A recent study showed that butyrate, a histone deacetylase (HDACs) suppresses the activation of human mast cells through the epigenetic regulation of the FcεRI-mediated pathway [234]. Phosphorylation of a 78kD protein or serine and threonine residues [235], which was later shown to be moesin [236], was shown to be associated with inhibition of secretion in rat mast cells [237] and basophils [238]. Primary mast cells from systemic mastocytosis patients are highly sensitive to suberoylanilide hydroxamic acid (SAHA)-induced cell death, whereas normal bone marrow mast cells are resistant. Thus, HDAC inhibitors may be a potential clinical treatment option for SM patients [239]. Histone acetylation is implicated in immune response associated with mast cell activation. Moreover, dietary substances could alter mast cell functions through epigenetic mechanisms [240]. An *in vitro* study demonstrated the role of epigenetics in the expression of CD34 and transcription factor HIF1A during the differentiation process of mast cells [241]. These reports indicate that epigenetic mechanisms could influence mast cell function and contribute to mast cell disorders.

6. Effect of neuropeptides

Most patients with MCADs present with neuropsychiatric symptoms and cognitive dysfunction [242–244]. The regulation of mast cells by neurotransmitters and neuropeptides has been reviewed [97,245,246], with emphasis on CRH [131], HK-1 [109], NGF [110], NT [132], SP [133] and somatostatin [134,135] acting via activation of high-affinity receptors.

Mast cells are also present in the central nervous system (CNS) [247,248], especially in the meninges [249,250], and the median eminence [249,251,252] where they are located perivascularly close to neurons [253] that are often positive for CRH secreted under stress [249]. Mast cells are the richest source of histamine in the CNS [254], particularly in the amygdala, hippocampus, hypothalamus and thalamus [255–257]. Brain mast cells had been termed the ‘immune gate to the brain’ [247]. Histamine is involved in neuronal development [258], and also acts as an alert signal in the brain [259]. In fact, brain histamine is critical for arousal [260,261], memory consolidation [262] and retrieval [263–265], as well as motivation [255]. Activated brain mast cells were shown to contribute to postoperative cognitive dysfunction via activation of microglia [266] involving mediators [267]. Cognitive dysfunction is common in mastocytosis patients [268,269] and may be related to morphological brain abnormalities, Primarily punctuated white matter abnormalities, that may potentially explain ‘brain fog.’

The effect of stress on mast cells has also been reviewed [102,270]. Restraint stress in rodents increased blood-brain barrier (BBB) permeability [251,271,272] via CRH-stimulating mast cells [271,273,274]. Mast cell-derived mediators, such as cytokines [275,276], increased BBB permeability not only to small molecules [251,271] but also to mammary adenocarcinoma brain metastases in mice [273]. This process could worsen with stress acting via CRH stimulation of mast cells [271,273], leading to increased dura vascular permeability, an effect that was absent in mast cell-deficient mice [277]. Interestingly, meningeal mast cells affected the integrity of the BBB and promoted T-cell brain infiltration [278].

Allergic stimulation of nasal mast cells resulted in stimulation of the hypothalamic-pituitary-adrenal (HPA) axis [131,279–281], possibly via mast cell release of histamine [282], IL-6 and CRH [108].

Mast cell responsiveness may be regulated not only by the neuroimmune stimuli but also by the effects of the different receptors involved. For instance, mast cells express high-affinity neurokinin-1 (NK-1) receptors for SP [210,283,284]. Moreover, SP [284] and NT [285] -induced the expression of CRHR-1 in human mast cells. Secretion of mediators can occur utilizing different signaling [20,221–223] and secretory [221,223] pathways. Interestingly, a circadian clock was reported to regulate IgE-dependent activation of murine bone marrow-derived mast cells [114,286,287].

7. Treatment approaches of mast cell-mediator-induced disorders

There are still no clinically effective inhibitors of mast cell activation (‘mast cell stabilizers’) [16,23,288,289]. Biologics have been used [17,290], mostly aiming at neutralizing the effect of IgE in urticaria with the use of omalizumab [291,292]. A number of inhibitors of tyrosine kinase (KIT) have been developed to block mast cell proliferation [293–295], but most of these drugs do not inhibit mast cell activation [296,297]. New approaches target the putative inhibitory receptor (Siglec-8) [17,298,299]. Disodium cromoglycate (cromolyn) is known as a ‘mast cell stabilizer’ because it had originally been shown to inhibit histamine release from rat peritoneal mast cells [237]. However, cromolyn does not effectively inhibit either mouse [300] or human [301] mast cells.

Minimizing exposure to potential triggers (e.g. allergens, foods, heat, stress) is clearly important. If there allergy to food antigens [302,303] or histamine intolerance is present [304], supplementation with the main histamine metabolizing enzyme, diamine oxidase (DAO) [305] shortly before meals could be beneficial especially if its activity is low or there is the presence of gene polymorphisms. The initial treatment approach is the use of second-generation, H-1 antihistamines up to 4 times the recommended doses as tolerated [306–310]. Unique among these, is the histamine-1 receptor antagonist rupatadine, which was specifically developed to have potent anti-PAF activity [311]. Rupatadine is not available in the US although it has been available in Europe for over 20 years and in Canada since about 2000 (Canadian online pharmacies will send it with a US prescription). Rupatadine at 40 mg/day is well tolerated and inhibits histamine- and PAF-induced flares and *ex vivo* platelet aggregation in normal male subjects [312]. When compared to other

second second-generation H-1 antihistamines in chronic urticaria, 20 mg/day of rupatadine showed the greatest efficacy in the treatment of CSU (71.6%) as compared to 20 mg/day of desloratadine (50%), and 20 mg/day of levocetirizine (21.7%) Notably, rupatadine also inhibited histamine and TNF release from human mast cells in response to PAF [256], and the release of histamine and IL-6 from human mast cells was stimulated by different triggers [313]. Rupatadine, unlike desloratadine and levocetirizine, also inhibited the PAF-induced release of histamine from human mast cells [314].

The first-generation H1 anti-histamine-1 ketotifen has been promoted as a mast cell inhibitor, but the evidence is weak [315,316] and it is associated with fatigue and weight gain.

Addition of H2- antihistamines may have additional benefits as cimetidine [317], ranitidine [318], and famotidine [319] have been reported to also inhibit rat mast cells. Interestingly human leukemic (HMC-1) and human skin mast cells primarily express H2 receptors [320].

Increasing evidence supports the premise that Vitamin D has immunoregulatory actions in general [321] and mast cell inhibitory functions in particular [322]. Specifically, vitamin D was reported to regulate IgE production [323] and inhibit IgE-dependent mast cell activation via specific vitamin D metabolites [324] and also inhibit ultraviolet B (UVB) irradiated skin mast cells [325]. Even though the precise mechanism is not known, the beneficial action of vitamin D may involve increased expression of the vitamin D receptor (VDR) receptor and inhibition of Lyn binding to FcεRI [326]. Vitamin D may also be important in inhibiting IL-33-associated mast cell activation as it was shown to enhance the production of soluble ST2 (IL-33sr), thus inhibiting the availability and action of IL-33 [327].

Certain naturally occurring flavonoids, commonly found in fruits, green plants and seeds, have potent antioxidant and anti-inflammatory properties [328,329]. Among these, the flavone luteolin stands out because it significantly inhibits the activation of both mast cells [301,330] and the brain macrophages, and microglia [331–334]. The novel luteolin structural analogue tetramethoxyluteolin (methoxyluteolin) is an even more potent inhibitor than luteolin [210,301,330,334] and has been incorporated into a skin lotion. Flavonoids have also been reported to modulate the synthesis of PAF [335], which has been strongly implicated in COVID-19 [336]. Luteolin especially formulated in olive pomace oil [337] to increase absorption may not only inhibit mast cell activation but also prevent neuroinflammation [338–341], is neuroprotective [338,342–344] and could reduce cognitive dysfunction [345–349], especially brain fog [350–352], the main complaint of many patients with MCADs [269,353]. However, data from controlled clinical trials are still lacking.

8. Conclusions

Mast cells play a critical role in atopic diseases [39], especially allergies [19] and anaphylactic reactions [2,39,354], but also in inflammation [2,44;202;204;205]. Symptoms associated with mast cell activation are numerous and can affect all organs making the presentation of patients confusing and diagnosis challenging. Symptoms may derive from numerous mediators released from mast cells other than histamine and tryptase necessitating a better

definition of mast cell activation. Early diagnosis and appropriate intervention, along with stress reduction, could minimize mast cell reactivity and improve quality of life.

9. Expert opinion

Great advances have been made in the potential treatment of the MCAD variant aggressive systemic mastocytosis.

There have been few advances in the development of better diagnostic panels for mast cell activation. Potential useful new serum biomarkers that would need to be validated could include CRH, eosinophil cationic protein (ECP), IL-6, MMP-9, neurotrophins, osteopontin [355,356], PAF, SP [357], as well as VEGF and other angiogenic factors [208,213,214,355,356,358]. Elevated serum levels of the nonspecific mast cell surface receptor MRGPRX2 may also be useful [359].

The ultimate goal should be better diagnosis and treatment of mast cell activation, especially in response to non-allergic triggers.

There should be directed funding toward better phenotyping of mast cells, especially in different conditions, developing useful biomarker panels for diagnosis of MCAS understanding the mechanism of selective mediator release without degranulation, and identifying ways to regulate mast cell reactivity and activation.

Critical for advancing the field is the availability of cultured primary human mast cells since the closest source so far is still immortalized leukemic mast cells [360] and since bone marrow-derived mast cells do not necessarily reflect the tissue mast cell phenotype [361].

Human organoids [362] with mast cells and other tissue cells (e.g. endothelial cells, fibroblasts, T cells, and brain cells) should be developed to mimic specific tissue environments and mast cell phenotype differentiation.

Brain mast cells should be studied as they may be involved in neurodevelopmental and neuroinflammatory conditions [76,245,363], especially since brain mast cells may not express FcεRI [364] but are able to release TNF [278] in response to other triggers.

Better research and education on aspects of mast cell activation-related diseases and their treatment, as well as increased communication among health professionals, is important in order to diagnose and treat MCADs [365].

Abbreviations

11β-PGF ₂ α	11β-Prostaglandin F ₂ α
BBB	Blood-brain barrier
CCL2	C-C motif chemokine ligand 2
RANTES	Regulated on Activation, Normal T Expressed and Secreted/CD117
	Cluster of differentiation 117
CNS	Central nervous system
COVID-19	Coronavirus disease of 2019
CPA3	Carboxypeptidase A3
CRH	Corticotropin-releasing hormone
CRHR-1	Corticotropin-releasing hormone receptor 1
CSU	Chronic spontaneous urticaria
CTMC	Connective tissue mast cells
CXCL8	C-X-C motif ligand 8
ECP	Eosinophil cationic protein

EDS	Ehlers Danlos syndrome
EMR2	=EGF-like module-containing mucin-like hormone receptor-like2
FcεRI	Fc epsilon receptor 1
HK-1	Hemokinin-1
HPA	Hypothalamic-pituitary-adrenal axis
IgE	Immunoglobulin E
IL-1β	Interleukin 1β
IBS	Irritable bowel syndrome
MCADs	Mast cell activation diseases
MCAS	Mast cell activation syndrome
MCAU	Mast cell activation unspecified
MCP-1	Monocyte chemoattractant protein 1
MCSS	Multiple chemical sensitivity syndrome
MMAS	Monoclonal Mast cell activation disorder
MMC	Mucosal mast cell
MMP	Matrix metalloproteinases
MMP-9	Matrix metalloproteinase-9
MRGPRX2	MAS-related G protein-coupled receptor-X2
NGF	Nerve growth factor
NK1	Neurokinin-1
NT	Neurotensin
PAF	Platelet-activating factor
PGD2	Prostaglandin D2
POTS	Postural orthostatic tachycardia syndrome
SARS-CoV-2	Severe acute respiratory syndrome coronavirus 2
SCF	Stem cell factor
Siglec-8	Sialic acid-binding immunoglobulin-like lectin 8
SP	Substance P
TNF	Tumor necrosis factor
VEGF	Vascular endothelial growth Factor

Declaration of interest

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References

Papers of special note have been highlighted as either of interest (*) or of considerable interest () to readers.**

1. Csaba G. Mast cell, the peculiar member of the immune system: a homeostatic aspect. *Acta Microbiol Immunol Hung.* **2015**;62(3):207–231.

- Theoharides TC, Cochrane DE. Critical role of mast cells in inflammatory diseases and the effect of acute stress. *J Neuroimmunol.* **2004**;146(1–2):1–12.
- Wernersson S, Pejler G. Mast cell secretory granules: armed for battle. *Nat Rev Immunol.* **2014**;14(7):478–494.
- Castells M. Mast cell mediators in allergic inflammation and mastocytosis. *Immunol Allergy Clin North Am.* **2006**;26(3):465–485.
- Mukai K, Tsai M, Saito H, et al. Mast cells as sources of cytokines, chemokines, and growth factors. *Immunol Rev.* **2018**;282(1):121–150. DOI:10.1111/imr.12634
- Csaba G, Forgacs A. The ontogenesis of mast cells. *Acta Biol Acad Sci Hung.* **1971**;22(4):423–430.
- Crivellato E, Travan L, Ribatti D. The phylogenetic profile of mast cells. *Methods Mol Biol.* **2015**;1220:11–27.
- Kapa E, Szigeti M, Juhasz A, et al. Phylogenesis of mast cells. I. Mast cells of the frog *Rana esculenta*. *Acta Biol Acad Sci Hung.* **1970**;21(2):141–147.
- Aldridge MC, Bismuth H. Gallbladder cancer: the polyp-cancer sequence. *Br J Surg.* **1990**;77(4):363–364.
- Mulero I, Sepulcre MP, Meseguer J, et al. Histamine is stored in mast cells of most evolutionarily advanced fish and regulates the fish inflammatory response. *Proc Natl Acad Sci U S A.* **2007**;104(49):19434–19439. DOI:10.1073/pnas.0704535104
- Crivellato E, Ribatti D, Mallardi F, et al. The mast cell: a multifunctional effector cell. *Adv Clin Path.* **2003**;7(1):13–26.
- Theoharides TC. The mast cell: a neuroimmunoendocrine master player. *Int J Tissue React.* **1996**;18(1):1–21.
- Parwaresch MR, Horny HP, Lennert K. Tissue mast cells in health and disease. *Pathol Res Pract.* **1985**;179(4–5):439–461.
- Siebenhaar F, Redegeld FA, Bischoff SC, et al. Mast cells as drivers of disease and therapeutic targets. *Trends Immunol.* **2018**;39:151–162.
- Falduto GH, Pfeiffer A, Luker A, et al. Emerging mechanisms contributing to mast cell-mediated pathophysiology with therapeutic implications. *Pharmacol Ther.* **2021**;220:107718.
- Dahlin JS, Maurer M, Metcalfe DD, et al. The ingenious mast cell: contemporary insights into mast cell behavior and function. *Allergy.* **2022**;77:83–99.
- Kolkhir P, Elieh-Ali-Komi D, Metz M, et al. Understanding human mast cells: lesson from therapies for allergic and non-allergic diseases. *Nat Rev Immunol.* **2022**;22(5):294–308. DOI:10.1038/s41577-021-00622-y
- Levi-Schaffer F, Gibbs BF, Hallgren J, et al. Selected recent advances in understanding the role of human mast cells in health and disease. *J Allergy Clin Immunol.* **2022**;149(6):1833–1844. DOI:10.1016/j.jaci.2022.01.030
- Olivera A, Beaven MA, Metcalfe DD. Mast cells signal their importance in health and disease. *J Allergy Clin Immunol.* **2018**;142(2):381–393.
- Sibilano R, Frossi B, Pucillo CE. Mast cell activation: a complex interplay of positive and negative signaling pathways. *Eur J Immunol.* **2014**;44(9):2558–2566.
- Gallenga CE, Pandolfi F, Caraffa A, et al. Interleukin-1 family cytokines and mast cells: activation and inhibition. *J Biol Regul Homeost Agents.* **2019**;33(1):1–6.
- Hakim-Rad K, Metz M, Maurer M. Mast cells: makers and breakers of allergic inflammation. *Curr Opin Allergy Clin Immunol.* **2009**;9(5):427–430.
- Theoharides TC, Alysandratos KD, Angelidou A, et al. Mast cells and inflammation. *Biochim Biophys Acta.* **2012**;1822(1):21–33. DOI:10.1016/j.bbadis.2010.12.014
- Kempuraj D, Ahmed ME, Selvakumar GP, et al. Brain Injury-Mediated Neuroinflammatory Response and Alzheimer's Disease. *Neuroscientist.* **2020**;26(2):134–155. DOI:10.1177/1073858419848293
- Ribatti D. The development of human mast cells. An historical reappraisal. *Exp Cell Res.* **2016**;342(2):210–215.
- Schmetzer O, Valentin P, Church MK, et al. Murine and human mast cell progenitors. *Eur J Pharmacol.* **2016**;778:2–10.
- Chen CC, Grimbaldston MA, Tsai M, et al. Identification of mast cell progenitors in adult mice. *Proc Natl Acad Sci U S A.* **2005**;102(32):11408–11413. DOI:10.1073/pnas.0504197102

28. Gurish MF, Austen KF. Developmental origin and functional specialization of mast cell subsets. *Immunity*. 2012;37:25–33.
29. Rivera J, Fierro NA, Olivera A, et al. New insights on mast cell activation via the high affinity receptor for IgE. *Adv Immunol*. 2008;98:85–120.
30. Rivera J, Gilfillan AM. Molecular regulation of mast cell activation. *J Allergy Clin Immunol*. 2006;117(6):1214–1225. quiz 26. DOI:10.1016/j.jaci.2006.04.015.
31. Hellman LT, Akula S, Thorpe M, et al. Tracing the origins of IgE, mast cells, and allergies by studies of wild animals. *Front Immunol*. 2017;8:1749.
32. Crivellato E, Nico B, Ribatti D. The history of the controversial relationship between mast cells and basophils. *Immunol Lett*. 2011;141(1):10–17.
33. Sa-Nunes A, Oliveira CJF, Ribeiro JM. Mast cells and basophils: from malevolent design to coevolutionary arms race. *Trends Parasitol*. 2020;36(8):655–659.
34. Cheng LE, Hartmann K, Roers A, et al. Perivascular mast cells dynamically probe cutaneous blood vessels to capture immunoglobulin E. *Immunity*. 2013;38(1):166–175. DOI:10.1016/j.immuni.2012.09.022
35. Peters JL, Cohen S, Staudenmayer J, et al. Prenatal negative life events increases cord blood IgE: interactions with dust mite allergen and maternal atopy. *Allergy*. 2012;67:545–551.
36. Msallam R, Balla J, Rathore APS, et al. Fetal mast cells mediate postnatal allergic responses dependent on maternal IgE. *Science*. 2020;370(6519):941–950. DOI:10.1126/science.aba0864
37. Lyons DO, Pullen NA. Beyond IgE: alternative mast cell activation across different disease states. *Int J Mol Sci* 2020; 21
- **This paper showed for the first time that fetal mast cells are mature enough to respond to maternal IgE and contribute to the development of atopic diseases in the to offspring.**
38. Luker AJ, Lownik JC, Conrad DH, et al. A new look at IgE beyond allergies. *F1000Res*. 2019;8:736.
39. Theoharides TC, Valent P, Akin C. Mast cells, mastocytosis, and related disorders. *N Engl J Med*. 2015;373(2):163–172.
40. Rodewald HR, Dessing M, Dvorak AM, et al. Identification of a committed precursor for the mast cell lineage. *Science*. 1996;271(5250):818–822. DOI:10.1126/science.271.5250.818
41. Kitamura Y, Ito A. Mast cell-committed progenitors. *Proc Natl Acad Sci U S A*. 2005;102(32):11129–11130.
42. Conti P, Pang X, Boucher W, et al. Impact of Rantes and MCP-1 chemokines on in vivo basophilic cell recruitment in rat skin injection model and their role in modifying the protein and mRNA levels for histidine decarboxylase. *Blood*. 1997;89:4120–4127.
43. Conti P, Reale M, Barbacone RC, et al. Intramuscular injection of hrRANTES causes mast cell recruitment and increased transcription of histidine decarboxylase in mice: lack of effects in genetically mast cell-deficient W/WV mice. *Faseb J*. 1998;12(15):1693–1700. DOI:10.1096/fasebj.12.15.1693
44. Galli SJ, Borregaard N, Wynn TA. Phenotypic and functional plasticity of cells of innate immunity: macrophages, mast cells and neutrophils. *Nat Immunol*. 2011;12(11):1035–1044.
45. Kitamura Y, Kanakura Y, Fujita J, et al. Differentiation and transdifferentiation of mast cells; a unique member of the hematopoietic cell family. *Int J Cell Cloning*. 1987;5(2):108–121. DOI:10.1002/stem.5530050203
46. Welker P, Grabbe J, Gibbs B, et al. Nerve growth factor- β induces mast-cell marker expression during in vitro culture of human umbilical cord blood cells. *Immunology*. 2000;99(3):418–426. DOI:10.1046/j.1365-2567.2000.00984.x
47. Leon A, Buriani A, Dal Toso R, et al. Mast cells synthesize, store, and release nerve growth factor. *Proc Natl Acad Sci U S A*. 1994;91(9):3739–3743. DOI:10.1073/pnas.91.9.3739
48. Theoharides TC. Skin mast cells: are we missing the forest for the trees? *Exp Dermatol*. 2016;25(6):422–423.
49. Welle M. Development, significance, and heterogeneity of mast cells with particular regard to the mast cell-specific proteases chymase and tryptase. *J Leukocyte Biol*. 1997;61(3):233–245.
50. Pejler G, Abrink M, Ringvall M, et al. Mast cell proteases. *Adv Immunol*. 2007;95:167–255.
51. Caughey GH. Serine proteinases of mast cell and leukocyte granules. A league of their own. *Am J Respir Crit Care Med*. 1994;150(6_pt_2):S138–42.
52. Tanaka S. Phenotypic and functional diversity of mast cells. *Int J Mol Sci*. 2020;21(11):3835.
53. McEuen AR, Gaca MD, Buckley MG, et al. Two distinct forms of human mast cell chymase—differences in affinity for heparin and in distribution in skin, heart, and other tissues. *Eur J Biochem*. 1998;256(2):461–470. DOI:10.1046/j.1432-1327.1998.2560461.x
54. Akula S, Hellman L, Aviles FX, et al. Analysis of the mast cell expressed carboxypeptidase A3 and its structural and evolutionary relationship to other vertebrate carboxypeptidases. *Dev Comp Immunol*. 2022;127:104273.
55. Atiakshin D, Kostin A, Trotsenko I, et al. Carboxypeptidase A3—A key component of the protease phenotype of mast cells. *Cells*. 2022;11(3):11. DOI:10.3390/cells11030570
56. Wang G, Fan WT, Zhang Z, et al. Expression of matrix metalloproteinase-8 and matrix metalloproteinase-13 in mast cells of human periapical lesions. *Int J Clin Exp Pathol*. 2018;11(5):2530–2536.
57. Xu L, Cai Z, Yang F, et al. Activation induced upregulation of MMP9 in mast cells is a positive feedback mediator for mast cell activation. *Mol Med Rep*. 2017;15(4):1759–1764. DOI:10.3892/mmr.2017.6215
58. Aborg CH, Bergendorff A, Bergqvist U, et al. Site of ionic binding of sodium and histamine in mast cell granules. *Br J Pharmacol*. 1968;34(1):195P–196P.
59. Frossi B, Mion F, Sibillano R, et al. Is it time for a new classification of mast cells? What do we know about mast cell heterogeneity? *Immunol Rev*. 2018;282(1):35–46. DOI:10.1111/immr.12636
60. Plum T, Wang X, Rettel M, et al. Human mast cell proteome reveals unique lineage, putative functions, and structural basis for cell ablation. *Immunity*. 2020;52(2):404–16 e5. DOI:10.1016/j.immuni.2020.01.012
61. Ronnberg E, Boey DZH, Ravindran A, et al. Immunoprofiling reveals novel mast cell receptors and the continuous nature of human lung mast cell heterogeneity. *Front Immunol*. 2022;12:804812.
62. Schofield JR, Afrin LB. Recognition and management of medication excipient reactivity in patients with mast cell activation syndrome. *Am J Med Sci*. 2019;357(6):507–511.
63. Horny HP, Valent P. Diagnosis of mastocytosis: general histopathological aspects, morphological criteria, and immunohistochemical findings. *Leuk Res*. 2001;25(7):543–551.
64. Valent P, Hartmann K, Bonadonna P, et al. Mast Cell Activation Syndromes: collegium Internationale Allergologicum Update 2022. *Int Arch Allergy Immunol*. 2022;183(7):693–705. DOI:10.1159/000524532.
- **An update on disorders involving mast cell activation sand information on overlapping clinical features, definitions, diagnostic criteria and personalized treatment.**
65. Killock D. A new standard for mastocytosis. *Nat Rev Clin Oncol*. 2022;19(2):71.
66. Shomali W, Gotlib J. Response criteria in advanced systemic mastocytosis: evolution in the Era of KIT inhibitors. *Int J Mol Sci*. 2021;22(6):2983.
67. Li Z. New insights into the pathogenesis of systemic mastocytosis. *Int J Mol Sci*. 2021;22(9):4900.
68. Valent P, Hartmann K, Bonadonna P, et al. Global classification of mast cell activation disorders: an ICD-10-CM-Adjusted proposal of the ECNM-AIM Consortium. *J Allergy Clin Immunol Pract*. 2022;10(8):1941–1950. DOI:10.1016/j.jaip.2022.05.007
69. Picard M, Giavina-Bianchi P, Mezzano V, et al. Expanding spectrum of mast cell activation disorders: monoclonal and idiopathic mast cell activation syndromes. *Clin Ther*. 2013;35(5):548–562. DOI:10.1016/j.clinthera.2013.04.001
70. Afrin LB, Self S, Menk J, et al. Characterization of mast cell activation syndrome. *Am J Med Sci*. 2017;353(3):207–215. DOI:10.1016/j.amjms.2016.12.013

71. Theoharides TC, Tsilioni I, Ren H. Recent advances in our understanding of mast cell activation – or should it be mast cell mediator disorders? *Expert Rev Clin Immunol*. 2019;15(6):639–656.
72. Afrin LB, Ackerley MB, Bluestein LS, et al. Diagnosis of mast cell activation syndrome: a global “consensus-2”. *Diagnosis (Berl)*. 2021;8:137–152.
73. Giannetti A, Filice E, Caffarelli C, et al. Mast cell activation disorders. *Med (Kaunas)*. 2021;57(2):124. DOI:10.3390/medicina57020124
74. Leru PM. Evaluation and classification of mast cell disorders: a difficult to manage pathology in clinical practice. *Cureus*. 2022;14:e22177.
75. Theoharides TC. Mast cells in irritable bowel syndrome and ulcerative colitis: function not numbers is what makes all the difference. *Dig Dis Sci*. 2014;59(5):897–898.
76. Theoharides TC, Kavalioti M, Tsilioni I. Mast cells, stress, fear and autism spectrum disorder. *Int J Mol Sci*. 2019;20(15):20.
77. Afrin LB. Some cases of hypermobile Ehlers–Danlos syndrome may be rooted in mast cell activation syndrome. *Am J Med Genet C Semin Med Genet*. 2021;187(4):466–472.
78. Kohno R, Cannom DS, Olshansky B, et al. Mast cell activation disorder and postural orthostatic tachycardia syndrome: a clinical association. *J Am Heart Assoc*. 2021;10(17):e021002. DOI:10.1161/JAHA.121.021002
79. Brock I, Prendergast W, Maitland A. Mast cell activation disease and immunoglobulin deficiency in patients with hypermobile Ehlers–Danlos syndrome/hypermobility spectrum disorder. *Am J Med Genet C Semin Med Genet*. 2021;187(4):473–481.
80. Kohn A, Chang C. The relationship between hypermobile Ehlers–Danlos syndrome (hEDS), postural orthostatic tachycardia syndrome (pots), and mast cell activation syndrome (MCAS). *Clin Rev Allergy Immunol*. 2020;58(3):273–297.
81. Novak P, Giannetti MP, Weller E, et al. Mast cell disorders are associated with decreased cerebral blood flow and small fiber neuropathy. *Annals of Allergy, Asthma & Immunol*. 2022;128(3):299–306 e1. DOI:10.1016/j.anai.2021.10.006
82. Eguiluz-Gracia I, Tay TR, Hew M, et al. Recent developments and highlights in biomarkers in allergic diseases and asthma. *Allergy*. 2018;73:2290–2305.
83. Ogulur I, Pat Y, Ardicli O, et al. Advances and highlights in biomarkers of allergic diseases. *Allergy*. 2021;76:3659–3686.
84. Commins SP. Diagnosis & management of alpha-gal syndrome: lessons from 2,500 patients. *Expert Rev Clin Immunol*. 2020;16(7):667–677.
85. Valent P, Akin CD. I think i am suffering from MCAS: differential diagnosis and separating facts from fiction. *J Allergy Clin Immunol Pract*. 2019;7(4):1109–1114.
86. Khokhar D, Akin C. Mast cell activation: when the whole is greater than the sum of its parts. *Med Clin North Am*. 2020;104(1):177–187.
87. Zhang S, Bernstein JA. Mast cell activation syndrome: myths and realities. *Allergy Asthma Proc*. 2021;42(3):198–204.
88. Jackson CW, Pratt CM, Rupprecht CP, et al. Mastocytosis and mast cell activation disorders: clearing the air. *Int J Mol Sci*. 2021;22(20):11270. DOI:10.3390/ijms22011270
89. Gulen T, Akin C, Bonadonna P, et al. Selecting the right criteria and proper classification to diagnose mast cell activation syndromes: a critical review. *J Allergy Clin Immunol Pract*. 2021;9(11):3918–3928. DOI:10.1016/j.jaip.2021.06.011
90. Buttgerit T, Gu S, Carneiro-Leao L, et al. Idiopathic mast cell activation syndrome is more often suspected than diagnosed—A prospective real-life study. *Allergy*. 2022;77(9):2794–2802. DOI:10.1111/all.15304
91. Akin C, Valent P, Metcalfe DD. Mast cell activation syndrome: proposed diagnostic criteria. *J Allergy Clin Immunol*. 2010;126(6):1099–104 e4.
92. Ravi A, Butterfield J, Weiler CR. Mast Cell Activation Syndrome: improved Identification by Combined Determinations of Serum Tryptase and 24-Hour Urine 11 β -Prostaglandin2 α . *J Allergy Clin Immunol Pract*. 2014;2(6):775–778.
93. Butterfield J, Weiler CR. The Utility of Measuring Urinary Metabolites of Mast Cell Mediators in Systemic Mastocytosis and Mast Cell Activation Syndrome. *J Allergy Clin Immunol Pract*. 2020;8(8):2533–2541. DOI:10.1016/j.jaip.2020.02.021.
- **This review highlights the importance of measuring different mast cell mediators.**
94. Giannetti MP, Godwin G, Weller E, et al. Differential mast cell mediators in systemic mastocytosis and hereditary α -tryptasemia. *J Allergy Clin Immunol*. 2022;150(5):1225–1227. DOI:10.1016/j.jaci.2022.04.025
95. Butterfield JH. Nontryptase Urinary and Hematologic Biomarkers of Mast Cell Expansion and Mast Cell Activation: status 2022. *J Allergy Clin Immunol Pract*. 2022;10(8):1974–1984.
96. Theoharides TC, Leeman SE. Effect of IL-33 on de novo synthesized mediators from human mast cells. *J Allergy Clin Immunol*. 2019;143(1):451.
97. Theoharides TC. Neuroendocrinology of mast cells: challenges and controversies. *Exp Dermatol*. 2017;26(9):751–759.
98. Theoharides TC. Need to define a subgroup of patients with idiopathic mast cell activation syndrome. *J Allergy Clin Immunol Pract*. 2022;10(4):1127–1128.
99. Ulambayar B, Yang EM, Cha HY, et al. Increased platelet activating factor levels in chronic spontaneous urticaria predicts refractoriness to antihistamine treatment: an observational study. *Clin Transl Allergy*. 2019;9(1):33. DOI:10.1186/s13601-019-0275-6
100. Upton JEM, Grunebaum E, Sussman G, et al. Platelet Activating Factor (PAF): a Mediator of Inflammation. *BioFactors*. 2022;48(6):1189–1202. DOI:10.1002/biof.1883
101. Vadas P, Perelman B, Liss G. Platelet-activating factor, histamine, and tryptase levels in human anaphylaxis. *J Allergy Clin Immunol*. 2013;131(1):144–149.
102. Theoharides TC. The impact of psychological stress on mast cells. *Annals of Allergy, Asthma & Immunol*. 2020;125(4):388–392.
103. Theoharides TC, Kempuraj D, Marchand J, et al. Urticaria pigmentosa associated with acute stress and lesional skin mast-cell expression of CRF-R1. *Clin Exp Dermatol*. 2009;34(5):e163–6. DOI:10.1111/j.1365-2230.2008.03043.x
104. Cao J, Papadopoulou N, Kempuraj D, et al. Human mast cells express corticotropin-releasing hormone (CRH) receptors and CRH leads to selective secretion of vascular endothelial growth factor. *J Immunol*. 2005;174(12):7665–7675. DOI:10.4049/jimmunol.174.12.7665
105. Theoharides TC, Stewart JM. Genitourinary mast cells and survival. *Transl Androl Urol*. *Transl Androl Urol*. 2015;4(5):579–586. DOI:10.3978/j.issn.2223-4683.2015.10.04.
- **This study showed that mast cells express functional CRH receptor and that CRH stimulates selective release VEGF from mast cells.**
106. Pang X, Cotreau-Bibbo MM, Sant GR, et al. Bladder mast cell expression of high affinity oestrogen receptors in patients with interstitial cystitis. *Br J Urol*. 1995;75(2):154–161. DOI:10.1111/j.1464-410X.1995.tb07303.x
107. Csaba G, Kovacs P. Hormones in the nucleus of mast cells: confocal microscopic immunocytochemical observations. *Horm Metab Res*. 2009;41(08):621–625.
108. Kempuraj D, Papadopoulou NG, Lytinas M, et al. Corticotropin-releasing hormone and its structurally related urocortin are synthesized and secreted by human mast cells. *Endocrinology*. 2004;145:43–48.
109. Sumpter TL, Ho CH, Pleet AR, et al. Autocrine hemokinin-1 functions as an endogenous adjuvant for IgE-mediated mast cell inflammatory responses. *J Allergy Clin Immunol*. 2015;135(4):1019–30 e8. DOI:10.1016/j.jaci.2014.07.036
110. Levi-Montalcini R, Skaper SD, Dal Toso R, et al. Nerve growth factor: from neurotrophin to neurokin. *Trends Neurosci*. 1996;19(11):514–520. DOI:10.1016/S0166-2236(96)10058-8
111. Hu H, Zhang R, Fang X, et al. Effects of endogenous substance P expression on degranulation in RBL-2H3 cells. *Inflamm Res*. 2011;60(6):541–546. DOI:10.1007/s00011-010-0301-6
112. Skaper SD, Pollock M, Facci L. Mast cells differentially express and release active high molecular weight neurotrophins. *Brain Res Mol Brain Res*. 2001;97(2):177–185.

113. Baumann A, Gonenwein S, Bischoff SC, et al. The circadian clock is functional in eosinophils and mast cells. *Immunology*. 2013;140:465–474.
114. Christ P, Sowa AS, Froy O, et al. The Circadian Clock Drives Mast Cell Functions in Allergic Reactions. *Front Immunol*. 2018;9:1526.
115. Pham L, Baiocchi L, Kennedy L, et al. The interplay between mast cells, pineal gland, and circadian rhythm: links between histamine, melatonin, and inflammatory mediators. *J Pineal Res*. 2021;70(2):e12699. DOI:10.1111/jpi.12699
116. Shimoi K, Okada H, Furugori M, et al. Intestinal absorption of luteolin and luteolin 7- O - β -glucoside in rats and humans. *FEBS Lett*. 1998;438(3):220–224. DOI:10.1016/S0014-5793(98)01304-0
117. Bachelet I, Munitz A, Moretta A, et al. The inhibitory receptor IRp60 (Cd300a) is expressed and functional on human mast cells. *J Immunol*. 2005;175(12):7989–7995. DOI:10.4049/jimmunol.175.12.7989
118. Karra L, Singh Gangwar R, Shamri R, et al. Leukocyte CD300a Contributes to the Resolution of Murine Allergic Inflammation. *J Immunol*. 2018;201(10):2998–3005. DOI:10.4049/jimmunol.1801000
119. Wang Y, Nakahashi-Oda C, Okayama Y, et al. Autonomous regulation of IgE-mediated mast cell degranulation and immediate hypersensitivity reaction by an inhibitory receptor CD300a. *J Allergy Clin Immunol*. 2019;144(1):323–7 e7. DOI:10.1016/j.jaci.2019.03.005
120. Vitalle J, Terren I, Orrantia A, et al. The Expression and Function of CD300 Molecules in the Main Players of Allergic Responses: mast Cells, Basophils and Eosinophils. *Int J Mol Sci*. 2020;21(9):3173. DOI:10.3390/ijms21093173
121. Smiljkovic D, Herrmann H, Sadovnik I, et al. Expression and regulation of Siglec-6 (CD327) on human mast cells and basophils. *J Allergy Clin Immunol*. 2023;151(1):202–211. DOI:10.1016/j.jaci.2022.07.018
122. Mizrahi S, Gibbs BF, Karra L, et al. Siglec-7 is an inhibitory receptor on human mast cells and basophils. *J Allergy Clin Immunol*. 2014;134(1):230–233. DOI:10.1016/j.jaci.2014.03.031
123. Landolina N, Zaffran I, Smiljkovic D, et al. Activation of Siglec-7 results in inhibition of in vitro and in vivo growth of human mast cell leukemia cells. *Pharmacol Res*. 2020;158:104682.
124. Altrichter S, Staubach P, Pasha M, et al. An open-label, proof-of-concept study of lirenlimab for antihistamine-resistant chronic spontaneous and inducible urticaria. *J Allergy Clin Immunol*. 2022;149(5):1683–90 e7. DOI:10.1016/j.jaci.2021.12.772
125. Bulfone-Paus S, Nilsson G, Draber P, et al. Positive and Negative Signals in Mast Cell Activation. *Trends Immunol*. 2017;38(9):657–667. DOI:10.1016/j.it.2017.01.008
126. Theoharides TC. Danger Signals and Inflammation. *Clin Ther*. 2016;38(5):996–999.
127. Redegeld FA, Yu Y, Kumari S, et al. Non-IgE mediated mast cell activation. *Immunol Rev*. 2018;282(1):87–113. DOI:10.1111/imr.12629
128. Migalovich-Sheikhet H, Friedman S, Mankuta D, et al. Novel identified receptors on mast cells. *Front Immunol*. 2012;3:238.
129. Elieh Ali Komi D, Shafaghat F, Kovanen PT, et al. Mast cells and complement system: ancient interactions between components of innate immunity. *Allergy*. 2020;75(11):2818–2828. DOI:10.1111/all.14413
130. Yanase Y, Takahagi S, Ozawa K, et al. The Role of Coagulation and Complement Factors for Mast Cell Activation in the Pathogenesis of Chronic Spontaneous Urticaria. *Cells*. 2021;10(7):1759. DOI:10.3390/cells10071759
131. Theoharides TC, Donelan JM, Papadopoulou N, et al. Mast cells as targets of corticotropin-releasing factor and related peptides. *Trends Pharmacol Sci*. 2004;25(11):563–568. DOI:10.1016/j.tips.2004.09.007
132. Donelan J, Boucher W, Papadopoulou N, et al. Corticotropin-releasing hormone induces skin vascular permeability through a neurotensin-dependent process. *Proc Natl Acad Sci U S A*. 2006;103(20):7759–7764. DOI:10.1073/pnas.0602210103
133. Theoharides TC, Zhang B, Kempuraj D, et al. IL-33 augments substance P-induced VEGF secretion from human mast cells and is increased in psoriatic skin. *Proc Natl Acad Sci U S A*. 2010;107(9):4448–4453. DOI:10.1073/pnas.1000803107.
- **Demonstrated that Substance P could activate human mast cells to release VEGF that was significantly increased with co-administration of IL-33.**
134. Theoharides TC, Betchaku T, Douglas WW. Somatostatin-induced histamine secretion in mast cells. Characterization of the effect. *Eur J Pharmacol*. 1981;69(2):127–137.
135. Theoharides TC, Douglas WW. Mast cell histamine secretion in response to somatostatin analogues: structural considerations. *Eur J Pharmacol*. 1981;73(2–3):131–136.
136. McNeil BD, Pundir P, Meeker S, et al. Identification of a mast-cell-specific receptor crucial for pseudo-allergic drug reactions. *Nature*. 2015;519:237–241.
137. Babina M, Wang Z, Li Z, et al. Fc ϵ RI- and MRGPRX2-evoked acute degranulation responses are fully additive in human skin mast cells. *Allergy*. 2022;77(6):1906–1909. DOI:10.1111/all.15270
138. Franke K, Wang Z, Zuberbier T, et al. Cytokines Stimulated by IL-33 in Human Skin Mast Cells: involvement of NF- κ B and p38 at Distinct Levels and Potent Co-Operation with Fc ϵ RI and MRGPRX2. *Int J Mol Sci*. 2021;22(7):3580. DOI:10.3390/ijms22073580
139. Sandig H, Bulfone-Paus S. TLR signaling in mast cells: common and unique features. *Front Immunol*. 2012;3:185.
140. Ogawa Y, Kinoshita M, Kawamura T, et al. Intracellular TLRs of Mast Cells in Innate and Acquired Immunity. *Handb Exp Pharmacol*. 2022;276:133–159.
141. Ratnaseelan AM, Tsilioni I, Theoharides TC. Effects of Mycotoxins on Neuropsychiatric Symptoms and Immune Processes. *Clin Ther*. 2018;40(6):903–917.
142. Marietta EV, Weis JJ, Weis JH. CD28 expression by mouse mast cells is modulated by lipopolysaccharide and outer surface protein a lipoprotein from *Borrelia burgdorferi*. *J Immunol*. 1997;159(6):2840–2848.
143. Bernard Q, Wang Z, Di Nardo A, et al. Interaction of primary mast cells with *Borrelia burgdorferi* (*sensu stricto*): role in transmission and dissemination in C57BL/6 mice. *Parasites Vectors*. 2017;10(1):313. DOI:10.1186/s13071-017-2243-0
144. Marshall JS, Portales-Cervantes L, Leong E. Mast Cell Responses to Viruses and Pathogen Products. *Int J Mol Sci*. 2019;20(17):20.
145. Abraham SN, St John AL. Mast cell-orchestrated immunity to pathogens. *Nat Rev Immunol*. 2010;10(6):440–452.
146. Song ST, Wu ML, Zhang HJ, et al. Mast Cell Activation Triggered by Retrovirus Promotes Acute Viral Infection. *Front Microbiol*. 2022;13:798660.
147. Afrin LB, Weinstock LB, Molderings GJ. Covid-19 hyperinflammation and post-Covid-19 illness may be rooted in mast cell activation syndrome. *Int J Infect Dis*. 2020;100:327–332.
148. Gebremeskel S, Schanin J, Coyle KM, et al. Mast Cell and Eosinophil Activation are Associated with COVID-19 and TLR-Mediated Viral Inflammation: implications for an Anti-Siglec-8 Antibody. *Front Immunol*. 2021;12:650331.
149. Motta Junior JDS, Miggiolaro A, Nagashima S, et al. Mast Cells in Alveolar Septa of COVID-19 Patients: a Pathogenic Pathway That May Link Interstitial Edema to Immuno-thrombosis. *Front Immunol*. 2020;11:574862.
150. Wu ML, Liu FL, Sun J, et al. SARS-CoV-2-triggered mast cell rapid degranulation induces alveolar epithelial inflammation and lung injury. *Signal Transduct Target Ther*. 2021;6(1): 428. DOI:10.1038/s41392-021-00849-0
151. Tan J, Anderson DE, Rathore APS, et al. Signatures of mast cell activation are associated with severe COVID-19. *medRxiv* 2021;
152. Theoharides TC. Potential association of mast cells with coronavirus disease 2019. *Annals of Allergy, Asthma & Immunol*. 2021;126(3):217–218.
153. Zelechowska P, Brzezinska-Blaszczczyk E, Agier J, et al. Different effectiveness of fungal pathogen-associated molecular patterns (PAMPs) in activating rat peritoneal mast cells. *Immunol Lett*. 2022;248:7–15.
154. Sayed BA, Brown MA. Mast cells as modulators of T-cell responses. *Immunol Rev*. 2007;217(1):53–64.

155. Krajewska NM, Fiancette R, Oo YH. Interplay between Mast Cells and Regulatory T Cells in Immune-Mediated Cholangiopathies. *Int J Mol Sci.* **2022**;23(11):5872.
156. Lotfi-Emran S, Ward BR, Le QT, et al. Human mast cells present antigen to autologous CD4(+) T cells. *J Allergy Clin Immunol.* **2018**;141(1):311–21 e10. DOI:10.1016/j.jaci.2017.02.048
157. Skokos D, Botros HG, Demeure C, et al. Mast cell-derived exosomes induce phenotypic and functional maturation of dendritic cells and elicit specific immune responses in vivo. *J Immunol.* **2003**;170(6):3037–3045. DOI:10.4049/jimmunol.170.6.3037
158. Schwartz LB. Mediators of human mast cells and human mast cell subsets. *Ann Allergy.* **1987**;58(4):226–235.
159. Uvnas B. Histamine storage and release. *Fed Proc.* **1974**;33(10):2172–2176.
160. Borriello F, Iannone R, Marone G. Histamine Release from Mast Cells and Basophils. *Handb Exp Pharmacol.* **2017**;241:121–139.
161. Cochrane DE, Carraway RE, Feldberg RS, et al. Stimulated rat mast cells generate histamine-releasing peptide from albumin. *Peptides.* **1993**;14:117–123.
162. Schwartz LB. Diagnostic value of tryptase in anaphylaxis and mastocytosis. *Immunol Allergy Clin North Am.* **2006**;26(3):451–463.
163. Bonadonna P, Scaffidi L, Boni E. Tryptase values in anaphylaxis and insect allergy. *Curr Opin Allergy Clin Immunol.* **2019**;19(5):462–467.
164. Valent P, Bonadonna P, Hartmann K, et al. Why the 20% + 2 Tryptase Formula is a Diagnostic Gold Standard for Severe Systemic Mast Cell Activation and Mast Cell Activation Syndrome. *Int Arch Allergy Immunol.* **2019**;180(1):44–51. DOI:10.1159/000501079
165. Fukuoka Y, Xia HZ, Sanchez-Munoz LB, et al. Generation of Anaphylatoxins by Human β -Tryptase from C3, C4, and C5. *J Immunol.* **2008**;180(9):6307–6316. DOI:10.4049/jimmunol.180.9.6307
166. Fiorucci L, Ascogli F. Mast cell tryptase, a still enigmatic enzyme. *Cell Mol Life Sci.* **2004**;61(11):1278–1295.
167. Awan SF, Schwartz LB, Maric I, et al. Acute increases in total serum tryptase unassociated with hemodynamic instability in diffuse cutaneous mastocytosis. *Annals of Allergy, Asthma & Immunol.* **2022**;129(2):249–252. DOI:10.1016/j.anai.2022.04.030
168. Boyce JA. Mast cells and eicosanoid mediators: a system of reciprocal paracrine and autocrine regulation. *Immunol Rev.* **2007**;217(1):168–185.
169. Varvara G, Tettamanti L, Gallenga CE, et al. Stimulated mast cells release inflammatory cytokines: potential suppression and therapeutic aspects. *J Biol Regul Homeost Agents.* **2018**;32(6):1355–1360.
170. Alevizos M, Karagkouni A, Vasiadi M, et al. Rupatadine inhibits inflammatory mediator release from human laboratory of allergic diseases 2 cultured mast cells stimulated by platelet-activating factor. *Annals of Allergy, Asthma & Immunol.* **2013**;111(6):542–547. DOI:10.1016/j.anai.2013.08.025
171. Demopoulos CA, Pinckard RN, Hanahan DJ. Platelet-activating factor. Evidence for 1-O-alkyl-2-acetyl-sn-glycerol-3-phosphorylcholine as the active component (a new class of lipid chemical mediators). *J Biol Chem.* **1979**;254(19):9355–9358.
172. Demopoulos C, Antonopoulou S, Theoharides TC. COVID-19, microthromboses, inflammation, and platelet activating factor. *BioFactors.* **2020**;46:927–933.
173. Theoharides TC, Antonopoulou S, Demopoulos CA. Coronavirus 2019, Microthromboses, and Platelet Activating Factor. *Clin Ther.* **2020**;42(10):1850–1852.
174. Demopoulos CA, Andrikopoulos NK, Antonopoulou S. A simple and precise method for the routine determination of platelet-activating factor in blood and urine. *Lipids.* **1994**;29:305–309.
175. Palgan K, Bartuzi Z. Platelet activating factor in allergies. *Int J Immunopathol Pharmacol.* **2015**;28(4):584–589.
176. Furukawa M, Ogura M, Tsutsumi T, et al. Presence of platelet-activating factor in nasal polyps and eosinophils. *Acta Otolaryngol.* **2002**;122(8):872–876. DOI:10.1080/003655402_000028064
177. Munoz-Cano RM, Casas-Saucedo R, Valero Santiago A, et al. Platelet-Activating Factor (PAF) in Allergic Rhinitis: clinical and Therapeutic Implications. *J Clin Med.* **2019**;8(9):1338. DOI:10.3390/jcm8091338
178. Austin CE, Foreman JC. The effect of platelet-activating factor on the responsiveness of the human nasal airway. *Br J Pharmacol.* **1993**;110(1):113–118.
179. Rihoux JP, Fadel R, Juhlin L. Platelet-activating factor-induced immediate and late cutaneous reactions. *Int Arch Allergy Appl Immunol.* **1991**;94(1–4):299–300.
180. Arias K, Baig M, Colangelo M, et al. Concurrent blockade of platelet-activating factor and histamine prevents life-threatening peanut-induced anaphylactic reactions. *J Allergy Clin Immunol.* **2009**;124(2):302–307. DOI:10.1016/j.jaci.2009.03.012
181. Tedeschi A, Palumbo G, Milazzo N, et al. Nasal neutrophilia and eosinophilia induced by challenge with platelet activating factor. *J Allergy Clin Immunol.* **1994**;93(2):526–533. DOI:10.1016/0091-6749(94)90363-8
182. Kato M, Kita H, Tachibana A, et al. Dual signaling and effector pathways mediate human eosinophil activation by platelet-activating factor. *Int Arch Allergy Immunol.* **2004**;Suppl 134(Suppl. 1):37–43. DOI:10.1159/000077791
183. Thivierge M, Rola-Pleszczynski M. Platelet-activating factor enhances interleukin-6 production by alveolar macrophages. *J Allergy Clin Immunol.* **1992**;90(5):796–802.
184. Keglowich L, Baraket M, Tamm M, et al. Hypoxia exerts dualistic effects on inflammatory and proliferative responses of healthy and asthmatic primary human bronchial smooth muscle cells. *PLoS ONE.* **2014**;9:e89875.
185. Hamel-Cote G, Lapointe F, Veronneau S, et al. Regulation of platelet-activating factor-mediated interleukin-6 promoter activation by the 48 kDa but not the 45 kDa isoform of protein tyrosine phosphatase non-receptor type 2. *Cell Biosci.* **2019**;9(1):51. DOI:10.1186/s13578-019-0316-9
186. Gutierrez S, Palacios I, Egidio J, et al. IL-1 β and IL-6 stimulate the production of platelet-activating factor (PAF) cultured rabbit synovial cells. *Clin Exp Immunol.* **1995**;99(3):364–368. DOI:10.1111/j.1365-2249.1995.tb05559.x
187. Biffi WL, Moore EE, Moore FA, et al. Interleukin-6 stimulates neutrophil production of platelet-activating factor. *J Leukocyte Biol.* **1996**;59(4):569–574. DOI:10.1002/jlb.59.4.569
188. Zhang B, Asadi S, Weng Z, et al. Stimulated human mast cells secrete mitochondrial components that have autocrine and paracrine inflammatory actions. *PLoS ONE.* **2012**;7:e49767.
189. Moon TC, Befus AD, Kulka M. Mast cell mediators: their differential release and the secretory pathways involved. *Front Immunol.* **2014**;5:569.
190. Theoharides TC, Bondy PK, Tsakalos ND, et al. Differential release of serotonin and histamine from mast cells. *Nature.* **1982**;297:229–231. DOI:10.1038/297229a0.
- **First reported that mast cells can release different mediators selectively without degranulation.**
191. Kandere-Grzybowska K, Letourneau R, Kempuraj D, et al. IL-1 induces vesicular secretion of IL-6 without degranulation from human mast cells. *J Immunol.* **2003**;171(9):4830–4836. DOI:10.4049/jimmunol.171.9.4830
192. Gagari E, Tsai M, Lantz CS, et al. Differential release of mast cell interleukin-6 via c-kit. *Blood.* **1997**;89:2654–2663.
193. Theoharides TC, Kempuraj D, Tagen M, et al. Differential release of mast cell mediators and the pathogenesis of inflammation. *Immunol Rev.* **2007**;217(1):65–78. DOI:10.1111/j.1600-065X.2007.00519.x
194. Theoharides TC, Douglas WW. Secretion in mast cells induced by calcium entrapped within phospholipid vesicles. *Science.* **1978**;201:1143–1145. DOI: 10.1126/science.684435.
- **First reported that mast cells can release the content of individual secretory granules without compound exocytosis.**
195. Dvorak AM. Piecemeal degranulation of basophils and mast cells is effected by vesicular transport of stored secretory granule contents. *Chem Immunol Allergy.* **2005**;85:135–184.
196. Crivellato E, Nico B, Gallo VP, et al. Cell secretion mediated by granule-associated vesicle transport: a glimpse at evolution. *Anat Rec.* **2010**;293(7):1115–1124. DOI:10.1002/ar.21146
197. Skokos D, Le Panse S, Villa I, et al. Mast cell-dependent B and T lymphocyte activation is mediated by the secretion of immunologically

- active exosomes. *J Immunol.* **2001**;166(2):868–876. DOI:10.4049/jimmunol.166.2.868
198. Dimitris S, Hany G-B, Michèle R, et al. Immunoregulatory properties of mast cell-derived exosomes. *Mol Immunol.* **2002**;38(16–18):1359–1362. DOI:10.1016/S0161-5890(02)00088-3
199. Shefler I, Salamon P, Hershko AY, et al. Mast cells as sources and targets of membrane vesicles. **2011**;17(34):3797–3804. DOI:10.2174/138161211798357836
200. Lecce M, Molfetta R, Milito ND, et al. FcεRI Signaling in the Modulation of Allergic Response: role of Mast Cell-Derived Exosomes. *Int J Mol Sci.* **2020**;21(15):5464. DOI:10.3390/ijms21155464
201. Shefler I, Salamon P, Mekori YA. Extracellular Vesicles as Emerging Players in Intercellular Communication: relevance in Mast Cell-Mediated Pathophysiology. *Int J Mol Sci.* **2021**;22(17):9176.
202. Phukan P, Barman B, Chengappa NK, et al. Diffusion tensor imaging analysis of rheumatoid arthritis patients with neuropsychiatric features to determine the alteration of white matter integrity due to vascular events. *Clin Rheumatol.* **2022**;41(10):3169–3177. DOI:10.1007/s10067-022-06262-4
203. Carroll-Portillo A, Surviladze Z, Cambi A, et al. Mast cell synapses and exosomes: membrane contacts for information exchange. *Front Immunol.* **2012**;3:46.
204. Joulia R, Gaudenzio N, Rodrigues M, et al. Mast cells form antibody-dependent degranulatory synapse for dedicated secretion and defence. *Nat Commun.* **2015**;6(1):6174. DOI:10.1038/ncomms7174
205. Weng Z, Zhang B, Tsilioni I, et al. Nanotube Formation: a Rapid Form of “Alarm Signaling”? *Clin Ther.* **2016**;38(5):1066–1072. DOI:10.1016/j.clinthera.2016.02.030
206. Enoksson M, Lyberg K, Moller-Westerberg C, et al. Mast cells as sensors of cell injury through IL-33 recognition. *J Immunol.* **2011**;186(4):2523–2528. DOI:10.4049/jimmunol.1003383
207. Saluja R, Khan M, Church MK, et al. The role of IL-33 and mast cells in allergy and inflammation. *Clin Transl Allergy.* **2015**;5(1):33. DOI:10.1186/s13601-015-0076-5
208. Cristinziano L, Poto R, Criscuolo G, et al. IL-33 and Superantigenic Activation of Human Lung Mast Cells Induce the Release of Angiogenic and Lymphangiogenic Factors. *Cells.* **2021**;10(1):145. DOI:10.3390/cells10010145
209. Petra AI, Tsilioni I, Taracanova A, et al. Interleukin 33 and interleukin 4 regulate interleukin 31 gene expression and secretion from human laboratory of allergic diseases 2 mast cells stimulated by substance P and/or immunoglobulin E. *Allergy Asthma Proc.* **2018**;39(2):153–160. DOI:10.2500/aap.2018.38.4105
210. Taracanova A, Alevizos M, Karagkouni A, et al. SP and IL-33 together markedly enhance TNF synthesis and secretion from human mast cells mediated by the interaction of their receptors. *Proc Natl Acad Sci U S A.* **2017**;114(20):E4002–9. DOI:10.1073/pnas.1524845114
- **Demonstrated that Substance P could activate human mast cells to release TNF that was significantly increased with co-administration of IL-33.**
211. Taracanova A, Tsilioni I, Conti P, et al. Substance P and IL-33 administered together stimulate a marked secretion of IL-1β from human mast cells, inhibited by methoxyluteolin. *Proc Natl Acad Sci U S A.* **2018**;115(40):E9381–90. DOI:10.1073/pnas.1810133115
212. Bawazeer MA, Theoharides TC. IL-33 stimulates human mast cell release of CCL5 and CCL2 via MAPK and NF-kappaB, inhibited by methoxyluteolin. *Eur J Pharmacol.* **2019**;865:172760.
213. Kay AB, Ying S, Ardelean E, et al. Calcitonin gene-related peptide and vascular endothelial growth factor are expressed in lesional but not uninvolved skin in chronic spontaneous urticaria. *Clin Exp Allergy.* **2014**;44(8):1053–1060. DOI:10.1111/cea.12348
214. Jagodzinska J, Polaniak R, Birkner E, et al. Analysis of circulating vascular endothelial growth factor and its soluble receptors in patients with different forms of chronic urticaria. *BioMed Res Int.* **2015**;2015:578383.
215. Conti P, Caraffa A, Tete G, et al. Mast cells activated by SARS-CoV-2 release histamine which increases IL-1 levels causing cytokine storm and inflammatory reaction in COVID-19. *J Biol Regul Homeost Agents.* **2020**;34(5):1629–1632. DOI:10.23812/20-2EDIT
216. Theoharides TC, Boucher W, Spear K. Serum interleukin-6 reflects disease severity and osteoporosis in mastocytosis patients. *Int Arch Allergy Immunol.* **2002**;128(4):344–350.
217. Brockow K, Akin C, Huber M, et al. IL-6 levels predict disease variant and extent of organ involvement in patients with mastocytosis. *Clin Immunol.* **2005**;115(2):216–223. DOI:10.1016/j.jclim.2005.01.011
218. Mayado A, Teodosio C, Garcia-Montero AC, et al. Increased IL6 plasma levels in indolent systemic mastocytosis patients are associated with high risk of disease progression. *Leukemia.* **2016**;30:124–130.
219. Kaur D, Gomez E, Doe C, et al. IL-33 drives airway hyper-responsiveness through IL-13-mediated mast cell: airway smooth muscle crosstalk. *Allergy.* **2015**;70:556–567.
220. Tobio A, Bandara G, Morris DA, et al. Oncogenic D816V-KIT signaling in mast cells causes persistent IL-6 production. *Haematologica.* **2020**;105:124–135.
221. Xu H, Bin NR, Sugita S. Diverse exocytic pathways for mast cell mediators. *Biochem Soc Trans.* **2018**;46(2):235–247.
222. Gilfillan AM, Tkaczyk C. Integrated signalling pathways for mast-cell activation. *Nat Rev Immunol.* **2006**;6(3):218–230.
223. Gaudenzio N, Sibillano R, Marichal T, et al. Different activation signals induce distinct mast cell degranulation strategies. *J Clin Invest.* **2016**;126(10):3981–3998. DOI:10.1172/JCI85538
224. Reszka E, Jablonska E, Wiczorek E, et al. Epigenetic Changes in Neoplastic Mast Cells and Potential Impact in Mastocytosis. *Int J Mol Sci.* **2021**;22(6):2964. DOI:10.3390/ijms22062964
225. Molderings GJ. Systemic mast cell activation disease variants and certain genetically determined comorbidities may be consequences of a common underlying epigenetic disease. *Med Hypotheses.* **2022**;163:110862.
226. Monticelli S, Leoni C. Epigenetic and transcriptional control of mast cell responses. *F1000Res.* **2017**;6:2064.
227. Molderings GJ. Transgenerational transmission of systemic mast cell activation disease-genetic and epigenetic features. *Transl Res.* **2016**;174:86–97.
228. Haenisch B, Frohlich H, Herms S, et al. Evidence for contribution of epigenetic mechanisms in the pathogenesis of systemic mast cell activation disease. *Immunogenetics.* **2014**;66:287–297. DOI:10.1007/s00251-014-0768-3.
- **Reported epigenetic processes in the pathogenesis of MCAD.**
229. Melo FR, Wallerman O, Paivandy A, et al. Tryptase-catalyzed core histone truncation: a novel epigenetic regulatory mechanism in mast cells. *J Allergy Clin Immunol.* **2017**;140(2):474–485. DOI:10.1016/j.jaci.2016.11.044
230. Alanazi S, Rabelo Melo F, Pejler G. Tryptase Regulates the Epigenetic Modification of Core Histones in Mast Cell Leukemia Cells. *Front Immunol.* **2021**;12:804408.
231. Leoni C, Montagner S, Rinaldi A, et al. Dnmt3a restrains mast cell inflammatory responses. *Proc Natl Acad Sci U S A.* **2017**;114(8):E1490–9. DOI:10.1073/pnas.1616420114
232. Alanazi S, Melo FR, Pejler G. Histone Methyltransferase Inhibition Has a Cytotoxic Impact on Transformed Mast Cells: implications for Mastocytosis. *Anticancer Res.* **2020**;40(5):2525–2536.
233. Rigo R, Chelbi R, Agopian J, et al. TET2 regulates immune tolerance in chronically activated mast cells. *JCI Insight.* **2022**;7. DOI:10.1172/jci.insight.154191.
234. Folkerts J, Redegeld F, Folkerts G, et al. Butyrate inhibits human mast cell activation via epigenetic regulation of FcεRI-mediated signaling. *Allergy.* **2020**;75(8):1966–1978. DOI:10.1111/all.14254
235. Correia I, Wang L, Pang X, et al. Characterization of the 78 kDa mast cell protein phosphorylated by the antiallergic drug cromolyn and homology to moesin. *Biochem Pharmacol.* **1996**;52(3):413–424. DOI:10.1016/0006-2952(96)00243-2
236. Theoharides TC, Wang L, Pang X, et al. Cloning and cellular localization of the rat mast cell 78-kDa protein phosphorylated in response to the mast cell stabilizer cromolyn. *J Pharmacol Exp Ther.* **2000**;294(3):810–821.
237. Theoharides TC, Sieghart W, Greengard P, et al. Antiallergic drug cromolyn may inhibit histamine secretion by regulating phosphorylation of a mast cell protein. *Science.* **1980**;207:80–82.

238. Olson FJ, Ludowyke RI, Karlsson NG. Discovery and identification of serine and threonine phosphorylated proteins in activated mast cells: implications for regulation of protein synthesis in the rat basophilic leukemia mast cell line RBL-2H3. *J Proteome Res.* 2009;8(6):3068–3077.
239. Lyberg K, Ali HA, Grootens J, et al. Histone deacetylase inhibitor SAHA mediates mast cell death and epigenetic silencing of constitutively active D816V KIT in systemic mastocytosis. *Oncotarget.* 2017;8:9647–9659.
240. Krajewski D, Kaczewski E, Rovatti J, et al. Epigenetic Regulation via Altered Histone Acetylation Results in Suppression of Mast Cell Function and Mast Cell-Mediated Food Allergic Responses. *Front Immunol.* 2018;9:2414.
241. Walczak-Drzewiecka A, Salkowska A, Ratajewski M, et al. Epigenetic regulation of CD34 and HIF1A expression during the differentiation of human mast cells. *Immunogenetics.* 2013;65:429–438.
242. Afrin LB, Pohlau D, Raithel M, et al. Mast cell activation disease: an underappreciated cause of neurological and psychiatric symptoms and diseases. *Brain Behav Immun.* 2015;50:314–321.
243. Jennings SV, Slee VM, Zack RM, et al. Patient Perceptions in Mast Cell Disorders. *Immunol Allergy Clin North Am.* 2018;38(3):505–525. DOI:10.1016/j.iac.2018.04.006
244. Russell N, Jennings S, Jennings B, et al. The Mastocytosis Society Survey on Mast Cell Disorders: part 2—Patient Clinical Experiences and Beyond. *J Allergy Clin Immunol Pract.* 2019;7(4):1157–65 e6. DOI:10.1016/j.jaip.2018.07.032
245. Theoharides TC, Tsilioni I, Bawazeer MMC. Neuroinflammation and Pain in Fibromyalgia Syndrome. *Front Cell Neurosci.* 2019;13:353.
246. Xu H, Shi X, Li X, et al. Neurotransmitter and neuropeptide regulation of mast cell function: a systematic review. *J Neuroinflammation.* 2020;17(1):356. DOI:10.1186/s12974-020-02029-3
247. Theoharides TC. Mast cells: the immune gate to the brain. *Life Sci.* 1990;46(9):607–617.
248. Traina G, Cocchi M. Mast cells in the brain – Old cells, new target. *J Integr Neurosci.* 2017;16(s1):S69–83.
249. Rozniecki JJ, Dimitriadou V, Lambracht-Hall M, et al. Morphological and functional demonstration of rat dura mater mast cell–neuron interactions in vitro and in vivo. *Brain Res.* 1999;849(1–2):1–15. DOI:10.1016/S0006-8993(99)01855-7
250. Polyzoidis S, Koletsis T, Panagiotidou S, et al. Mast cells in meningiomas and brain inflammation. *J Neuroinflammation.* 2015;12(1):170. DOI:10.1186/s12974-015-0388-3
251. Theoharides TC. Corticotropin-releasing hormone and the blood-brain-barrier. *Front Biosci.* 2007;12(1):1615–1628.
252. Colonna M, Butovsky O. Microglia Function in the Central Nervous System During Health and Neurodegeneration. *Annu Rev Immunol.* 2017;35(1):441–468.
253. Paus R, Theoharides TC, Arck PC. Neuroimmunoendocrine circuitry of the 'brain-skin connection'. *Trends Immunol.* 2006;27(1):32–39.
254. Panula P. Histamine receptors, agonists, and antagonists in health and disease. *Handb Clin Neurol.* 2021;180:377–387.
255. Torrealba F, Riveros ME, Contreras M, et al. Histamine and motivation. *Front Syst Neurosci.* 2012;6:51.
256. Nomura H, Shimizume R, Ikegaya Y. Histamine: a Key Neuromodulator of Memory Consolidation and Retrieval. *Curr Top Behav Neurosci.* 2021;59:329–353.
257. Theoharides TC. Mast cells in atopic diseases: more than just histamine. eds., Jain V HN, Ohayon J, Wasserman S., Ohayon J. 2022.
258. Carthy E, Ellender TH. Neuroinflammation and Neurodevelopment: a Review. *Front Neurosci.* 2021;15:680214.
259. Mochizuki T. Histamine as an Alert Signal in the Brain. *Curr Top Behav Neurosci.* 2021;59:413–425.
260. Sadek B, Saad A, Sadeq A, et al. Histamine H3 receptor as a potential target for cognitive symptoms in neuropsychiatric diseases. *Behav Brain Res.* 2016;312:415–430.
261. Schlicker E, Kathmann M. Role of the Histamine H3 Receptor in the Central Nervous System. *Handb Exp Pharmacol.* 2017;241:277–299.
262. da Silveira CK, Furini CR, Benetti F, et al. The role of histamine receptors in the consolidation of object recognition memory. *Neurobiol Learn Mem.* 2013;103:64–71.
263. Provensi G, Costa A, Izquierdo I, et al. Brain histamine modulates recognition memory: possible implications in major cognitive disorders. *Br J Pharmacol.* 2020;177(3):539–556. DOI:10.1111/bph.14478
264. Passani MB, Benetti F, Blandina P, et al. Histamine regulates memory consolidation. *Neurobiol Learn Mem.* 2017;145:1–6.
265. Burgess CR. Histamine and orexin in the control of arousal, locomotion, and motivation. *J Neurosci.* 2010;30(8):2810–2811.
266. Zhang X, Dong H, Li N, et al. Activated brain mast cells contribute to postoperative cognitive dysfunction by evoking microglia activation and neuronal apoptosis. *J Neuroinflammation.* 2016;13(1):127. DOI:10.1186/s12974-016-0592-9.
- **This study showed that mast cell activation leads to the activation of microglia and exacerbates neuroinflammation.**
267. Zhang X, Wang Y, Dong H, et al. Induction of Microglial Activation by Mediators Released from Mast Cells. *Cell Physiol Biochem.* 2016;38(4):1520–1531. DOI:10.1159/000443093
268. Spolak-Bobryk N, Romantowski J, Kujawska-Danecka H, et al. Mastocytosis patients' cognitive dysfunctions correlate with the presence of spindle-shaped mast cells in bone marrow. *Clin Transl Allergy.* 2022;12(1):e12093. DOI:10.1002/ct2.12093
269. Moura DS, Sultan S, Georjin-Lavialle S, et al. Evidence for cognitive impairment in mastocytosis: prevalence, features and correlations to depression. *PLoS ONE.* 2012;7:e39468.
270. Theoharides TC. Effect of Stress on Neuroimmune Processes. *Clin Ther.* 2020;42(6):1007–1014.
271. Esposito P, Chandler N, Kandere K, et al. Corticotropin-releasing hormone and brain mast cells regulate blood-brain-barrier permeability induced by acute stress. *J Pharmacol Exp Ther.* 2002;303(3):1061–1066. DOI:10.1124/jpet.102.038497
272. Fiorentino M, Sapone A, Senger S, et al. Blood–brain barrier and intestinal epithelial barrier alterations in autism spectrum disorders. *Mol Autism.* 2016;7(1):49. DOI:10.1186/s13229-016-0110-z
273. Rozniecki JJ, Sahagian GG, Kempuraj D, et al. Brain metastases of mouse mammary adenocarcinoma is increased by acute stress. *Brain Res.* 2010;1366:204–210.
274. Theoharides TC, Rozniecki JJ, Sahagian G, et al. Impact of stress and mast cells on brain metastases. *J Neuroimmunol.* 2008;205(1–2):1–7. DOI:10.1016/j.jneuroim.2008.09.014
275. Abbott NJ. Inflammatory mediators and modulation of blood-brain barrier permeability. *Cell Mol Neurobiol.* 2000;20(2):131–147.
276. Pan W, Stone KP, Hsueh H, et al. Cytokine signaling modulates blood-brain barrier function. 2011;17(33):3729–3740. DOI:10.2174/138161211798220918
277. Kandere-Grzybowska K, Gheorghie D, Priller J, et al. Stress-induced dura vascular permeability does not develop in mast cell-deficient and neurokinin-1 receptor knockout mice. *Brain Res.* 2003;980(2):213–220. DOI:10.1016/S0006-8993(03)02975-5
278. Sayed BA, Christy AL, Walker ME, et al. Meningeal mast cells affect early T cell central nervous system infiltration and blood-brain barrier integrity through TNF: a role for neutrophil recruitment? *J Immunol.* 2010;184(12):6891–6900. DOI:10.4049/jimmunol.1000126
279. Matsumoto I, Inoue Y, Shimada T, et al. Brain mast cells act as an immune gate to the hypothalamic-pituitary-adrenal axis in dogs. *J Exp Med.* 2001;194(1):71–78. DOI:10.1084/jem.194.1.71
280. Bugajski AJ, Chlap Z, Gadek-Michalska A, et al. Degranulation and decrease in histamine levels of thalamic mast cells coincides with corticosterone secretion induced by compound 48/80. *Inflamm Res.* 1995;44(Suppl S1):S50–1. DOI:10.1007/BF01674391
281. Kalogeromitros D, Syrigou EK, Makris M, et al. Nasal provocation of patients with allergic rhinitis and the hypothalamic-pituitary-adrenal axis. *Annals of Allergy, Asthma & Immunol.* 2007;98(3):269–273. DOI:10.1016/S1081-1206(10)60717-X
282. Scaccianoce S, Lombardo K, Nicolai R, et al. Studies on the involvement of histamine in the hypothalamic-pituitary-adrenal axis activation induced by nerve growth factor. *Life Sci.* 2000;67(26):3143–3152. DOI:10.1016/S0024-3205(00)00899-7
283. Okada T, Hirayama Y, Kishi S, et al. Functional neurokinin NK-1 receptor expression in rat peritoneal mast cells. *Inflamm Res.* 1999;48(5):274–279. DOI:10.1007/s000110050459

284. Asadi S, Alysandratos KD, Angelidou A, et al. Substance P (SP) induces expression of functional corticotropin-releasing hormone receptor-1 (CRHR-1) in human mast cells. *J Invest Dermatol*. 2012;132(2):324–329. DOI:10.1038/jid.2011.334
285. Alysandratos KD, Asadi S, Angelidou A, et al. Neurotensin and CRH interactions augment human mast cell activation. *PLoS ONE*. 2012;7:e48934.
286. Wang X, Reece SP, Van Scott MR, et al. A circadian clock in murine bone marrow-derived mast cells modulates IgE-dependent activation in vitro. *Brain Behav Immun*. 2011;25(1):127–134. DOI:10.1016/j.bbi.2010.09.007
287. Kawauchi T, Ishimaru K, Nakamura Y, et al. Clock-dependent temporal regulation of IL-33/ST2-mediated mast cell response. *Allergol Int*. 2017;66(3):472–478. DOI:10.1016/j.alit.2017.02.004
288. Finn DF, Walsh JJ. Twenty-first century mast cell stabilizers. *Br J Pharmacol*. 2013;170(1):23–37.
289. Burchett JR, Dailey JM, Kee SA, et al. Targeting Mast Cells in Allergic Disease: current Therapies and Drug Repurposing. *Cells*. 2022;11(19):11. DOI:10.3390/cells11193031
290. Lyons JJ, Metcalfe DD. Targeting Mast Cells with Biologics. *Immunol Allergy Clin North Am*. 2020;40(4):667–685.
291. Maurer M, Metz M, Brehler R, et al. Omalizumab treatment in patients with chronic inducible urticaria: a systematic review of published evidence. *J Allergy Clin Immunol*. 2018;141(2):638–649. DOI:10.1016/j.jaci.2017.06.032
292. Ando T, Kitaura J. Tuning IgE: igE-Associating Molecules and Their Effects on IgE-Dependent Mast Cell Reactions. *Cells*. 2021;10(7):1697.
293. Giannetti MP. Treatment of systemic mastocytosis: novel and emerging therapies. *Annals of Allergy, Asthma & Immunol*. 2021;127(4):412–419.
294. Caslin HL, Kiwanuka KN, Haque TT, et al. Controlling Mast Cell Activation and Homeostasis: work Influenced by Bill Paul That Continues Today. *Front Immunol*. 2018;9:868.
295. Heinrich MC, Griffith DJ, Druker BJ, et al. Inhibition of c-kit receptor tyrosine kinase activity by STI 571, a selective tyrosine kinase inhibitor. *Blood*. 2000;96:925–932.
296. Gotlib J, Kluijn-Nelemans HC, George TI, et al. Efficacy and Safety of Midostaurin in Advanced Systemic Mastocytosis. *N Engl J Med*. 2016;374(26):2530–2541. DOI:10.1056/NEJMoa1513098
297. Akin C, Arock M, Valent P. Tyrosine kinase inhibitors for the treatment of indolent systemic mastocytosis: are we there yet? *J Allergy Clin Immunol*. 2022;149(6):1912–1918.
298. Duguay BA, Lu L, Arizmendi N, et al. The Possible Uses and Challenges of Nanomaterials in Mast Cell Research. *J Immunol*. 2020;204(8):2021–2032. DOI:10.4049/jimmunol.1800658
299. Duan S, Arlian BM, Nycholat CM, et al. Nanoparticles Displaying Allergen and Siglec-8 Ligands Suppress IgE-FcεRI-Mediated Anaphylaxis and Desensitize Mast Cells to Subsequent Antigen Challenge. *J Immunol*. 2021;206(10):2290–2300. DOI:10.4049/jimmunol.1901212
300. Oka T, Kalesnikoff J, Starkl P, et al. Evidence questioning cromolyn's effectiveness and selectivity as a 'mast cell stabilizer' in mice. *Lab Invest*. 2012;92(10):1472–1482. DOI:10.1038/labinvest.2012.116
301. Weng Z, Patel AB, Panagiotidou S, et al. The novel flavone tetramethoxyluteolin is a potent inhibitor of human mast cells. *J Allergy Clin Immunol*. 2015;135(4):1044–52 e5. DOI:10.1016/j.jaci.2014.10.032
302. Aguilera-Lizarraga J, Florens MV, Viola MF, et al. Local immune response to food antigens drives meal-induced abdominal pain. *Nature*. 2021;590:151–156.
303. Rothenberg ME, Phimister EG. An Allergic Basis for Abdominal Pain. *N Engl J Med*. 2021;384(22):2156–2158.
304. Maintz L, Novak N. Histamine and histamine intolerance. *Am J Clin Nutr*. 2007;85(5):1185–1196.
305. Podder I, Jaiswal S, Das A. Dietary strategies for chronic spontaneous urticaria: an evidence-based review. *Int J Dermatol*. 2021;62(2):143–153.
306. Shi Y, Zhou S, Zheng Q, et al. Systematic reviews of pharmacological and nonpharmacological treatments for patients with chronic urticaria: an umbrella systematic review. *Medicine (Baltimore)*. 2019;98:e15711.
307. Yanai K, Yoshikawa T, Church MK. Efficacy and Safety of Non-brain Penetrating H1-Antihistamines for the Treatment of Allergic Diseases. *Curr Top Behav Neurosci*. 2021;59:193–214.
308. Iriarte Sotes P, Armisen M, Usero-Barcelona T, et al. Efficacy and Safety of Up-dosing Antihistamines in Chronic Spontaneous Urticaria: a Systematic Review of the Literature. *J Investig Allergol Clin Immunol*. 2021;31(4):282–291. DOI:10.18176/jiaci.0649
309. Sarti L, Barni S, Giovannini M, et al. Efficacy and tolerability of the up-dosing of second-generation non-sedating H1 antihistamines in children with chronic spontaneous urticaria. *Pediatr Allergy Immunol*. 2021;32(1):153–160. DOI:10.1111/pai.13325
310. He L, Yi W, Huang X, et al. Chronic Urticaria: advances in Understanding of the Disease and Clinical Management. *Clin Rev Allergy Immunol*. 2021;61(3):424–448. DOI:10.1007/s12016-021-08886-x
311. Gonzalez-Nunez V, Bachert C, Mullol J. Rupatadine: global safety evaluation in allergic rhinitis and urticaria. *Expert Opin Drug Saf*. 2016;15(10):1439–1448.
312. Church MK. Efficacy and tolerability of rupatadine at four times the recommended dose against histamine- and platelet-activating factor-induced flare responses and ex vivo platelet aggregation in healthy males. *Br J Dermatol*. 2010;163(6):1330–1332.
313. Vasiadi M, Kalogeromitros D, Kempuraj D, et al. Rupatadine inhibits proinflammatory mediator secretion from human mast cells triggered by different stimuli. *Int Arch Allergy Immunol*. 2010;151(1):38–45. DOI:10.1159/000232569
314. Munoz-Cano R, Ainsua-Enrich E, Torres-Atencio I, et al. Effects of Rupatadine on Platelet-Activating Factor-Induced Human Mast Cell Degranulation Compared with Desloratadine and Levocetirizine (The MASPAF Study). *J Investig Allergol Clin Immunol*. 2017;27(3):161–168. DOI:10.18176/jiaci.0117
315. Huston DP, Bressler RB, Kaliner M, et al. Prevention of mast-cell degranulation by ketotifen in patients with physical urticarias. *Ann Intern Med*. 1986;104(4):507–510. DOI:10.7326/0003-4819-104-4-507
316. Schoch C. In vitro inhibition of human conjunctival mast-cell degranulation by ketotifen. *J Ocul Pharmacol Ther*. 2003;19(1):75–81.
317. He SH, Xie H, Fu YL. Inhibition of tryptase release from human colon mast cells by histamine receptor antagonists. *Asian Pac J Allergy Immunol*. 2005;23(1):35–39.
318. Lippert U, Moller A, Welker P, et al. Inhibition of cytokine secretion from human leukemic mast cells and basophils by H1- and H2-receptor antagonists. *Exp Dermatol*. 2000;9(2):118–124. DOI:10.1034/j.1600-0625.2000.09002118.x
319. Shah PM, Boulos PB, Springall R, et al. Effects of the H2-antagonists famotidine and nizatidine and the cytoprotectant misoprostol on human colonic and rat peritoneal mast cells. *Agents Actions*. 1994;41(S1):C51–52. Spec No:C51-2. DOI:10.1007/BF02007763.
320. Lippert U, Artuc M, Grutzkau A, et al. Human skin mast cells express H2 and H4, but not H3 receptors. *J Invest Dermatol*. 2004;123(1):116–123. DOI:10.1111/j.0022-202X.2004.22721.x
321. Theoharides TC. Vitamin D and Atopy. *Clin Ther*. 2017;39:880–883.
322. Murdaca G, Allegra A, Tonacci A, et al. Mast Cells and Vitamin D Status: a Clinical and Biological Link in the Onset of Allergy and Bone Diseases. *Biomedicines*. 2022;10(8):1877.
323. James J, Weaver V, Cantorna MT. Control of Circulating IgE by the Vitamin D Receptor in vivo Involves B Cell Intrinsic and Extrinsic Mechanisms. *J Immunol*. 2017;198:1164–1171.
324. Yip KH, Kolesnikoff N, Yu C, et al. Mechanisms of vitamin D(3) metabolite repression of IgE-dependent mast cell activation. *J Allergy Clin Immunol*. 2014;133:1356–1364.
325. Yu C, Fedoric B, Anderson PH, et al. Vitamin D(3) signalling to mast cells: a new regulatory axis. *Int J Biochem Cell Biol*. 2011;43:41–46.
326. Liu ZQ, Li XX, Qiu SQ, et al. Vitamin D contributes to mast cell stabilization. *Allergy*. 2017;72:1184–1192.
327. Pfeffer PE, Chen YH, Woszczek G, et al. Vitamin D enhances production of soluble ST2, inhibiting the action of IL-33. *J Allergy Clin Immunol*. 2015;135:824–7 e3.
328. Middleton E Jr., Kandaswami C, Theoharides TC. The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease, and cancer. *Pharmacol Rev*. 2000;52:673–751.

329. Al-Khayri JM, Sahana GR, Nagella P, et al. Flavonoids as Potential Anti-Inflammatory Molecules: a Review. *Molecules*. 2022;27(9):2901.
330. Patel AB, Theoharides TC. Methoxyluteolin Inhibits Neuropeptide-stimulated Proinflammatory Mediator Release via mTOR Activation from Human Mast Cells. *J Pharmacol Exp Ther*. 2017;361:462–471.
331. Rezai-Zadeh K, Ehrhart J, Bai Y, et al. Apigenin and luteolin modulate microglial activation via inhibition of STAT1-induced CD40 expression. *J Neuroinflammation*. 2008;5:41.
332. Jang S, Kelley KW, Johnson RW. Luteolin reduces IL-6 production in microglia by inhibiting JNK phosphorylation and activation of AP-1. *Proc Natl Acad Sci U S A*. 2008;105:7534–7539.
333. Burton MD, Rytych JL, Amin R, et al. Dietary Luteolin Reduces Proinflammatory Microglia in the Brain of Senescent Mice. *Rejuvenation Res*. 2016;19:286–292.
334. Patel AB, Tsiloni I, Leeman SE, et al. Neurotensin stimulates sortilin and mTOR in human microglia inhibitable by methoxyluteolin, a potential therapeutic target for autism. *Proc Natl Acad Sci U S A*. 2016;113:E7049–58.
335. Balestrieri ML, Castaldo D, Balestrieri C, et al. Modulation by flavonoids of PAF and related phospholipids in endothelial cells during oxidative stress. *J Lipid Res*. 2003;44:380–387.
336. Klein M, Dao V, Khan F. A Review of Platelet-Activating Factor as a Potential Contributor to Morbidity and Mortality Associated with Severe COVID-19. *Clin Appl Thromb Hemost*. 2021;27:10760296211051764.
337. Theoharides TC. Luteolin supplements: all that glitters is not gold. *BioFactors*. 2021;47:242–244.
338. Ashaari Z, Hadjzadeh MA, Hassanzadeh G, et al. The Flavone Luteolin Improves Central Nervous System Disorders by Different Mechanisms: a Review. *J Mol Neurosci*. 2018;65:491–506.
339. Calis Z, Mogulkoc R, Baltaci AK. The Roles of Flavonols/Flavonoids in Neurodegeneration and Neuroinflammation. *Mini Rev Med Chem*. 2020;20:1475–1488.
340. Kempuraj D, Thangavel R, Kempuraj DD, et al. Neuroprotective effects of flavone luteolin in neuroinflammation and neurotrauma. *BioFactors*. 2021;47:190–197.
341. Theoharides TC, Conti P, Economu M. Brain inflammation, neuropsychiatric disorders, and immunoendocrine effects of luteolin. *J Clin Psychopharmacol*. 2014;34:187–189.
342. Khalili N, Haseli S, Bahrami-Motlagh H, et al. Neurologic Involvement in COVID-19: radiologists. *Perspective Acad Radiol*. 2020;27(7):1051.
343. Dajas F, Rivera-Megret F, Blasina F, et al. Neuroprotection by flavonoids. *Braz J Med Biol Res*. 2003;36:1613–1620.
344. Lin TY, Lu CW, Wang SJ. Luteolin protects the hippocampus against neuron impairments induced by kainic acid in rats. *Neurotoxicology*. 2016;55:48–57.
345. Rezai-Zadeh K, Douglas Shytle R, Bai Y, et al. Flavonoid-mediated presenilin-1 phosphorylation reduces Alzheimer's disease beta-amyloid production. *J Cell Mol Med*. 2009;13:574–588.
346. Theoharides TC, Stewart JM, Hatziaelaki E, et al. Brain “fog,” inflammation and obesity: key aspects of neuropsychiatric disorders improved by luteolin. *Front Neurosci*. 2015;9:225.
347. Yao ZH, Yao XL, Zhang Y, et al. Luteolin Could Improve Cognitive Dysfunction by Inhibiting Neuroinflammation. *Neurochem Res*. 2018;43:806–820.
348. Gratton G, Weaver SR, Burley CV, et al. Dietary flavanols improve cerebral cortical oxygenation and cognition in healthy adults. *Sci Rep*. 2020;10:19409.
349. Devi SA, Chamoli A. Polyphenols as an Effective Therapeutic Intervention Against Cognitive Decline During Normal and Pathological Brain Aging. *Adv Exp Med Biol*. 2020;1260:159–174.
350. Theoharides TC, Cholevas C, Polyzoidis K, et al. Long-COVID syndrome-associated brain fog and chemofog: luteolin to the rescue. *BioFactors*. 2021;47:232–241.
- **Reported that the flavonoid luteolin can prevent the development of neuropsychiatric disorders in Long-COVID.**
351. Stefano GB, Buttiker P, Weissenberger S, et al. Editorial: the Pathogenesis of Long-Term Neuropsychiatric COVID-19 and the Role of Microglia, Mitochondria, and Persistent Neuroinflammation: a Hypothesis. *Med Sci Monit*. 2021;27:e933015.
352. Hugon J, Msika EF, Queneau M, et al. Long COVID: cognitive complaints (brain fog) and dysfunction of the cingulate cortex. *J Neurol*. 2022;269:44–46.
353. Jennings S, Russell N, Jennings B, et al. The Mastocytosis Society survey on mast cell disorders: patient experiences and perceptions. *J Allergy Clin Immunol Pract*. 2014;2:70–76.
354. Bachelet I, Levi-Schaffer F, Mekori YA. Mast cells: not only in allergy. *Immunol Allergy Clin North Am*. 2006;26:407–425.
355. Bulfone-Paus S, Paus R. Osteopontin as a new player in mast cell biology. *Eur J Immunol*. 2008;38:338–341.
356. Nagasaka A, Matsue H, Matsushima H, et al. Osteopontin is produced by mast cells and affects IgE-mediated degranulation and migration of mast cells. *Eur J Immunol*. 2008;38:489–499.
357. Peng WM, Maintz L, Allam JP, et al. Increased circulating levels of neurotrophins and elevated expression of their high-affinity receptors on skin and gut mast cells in mastocytosis. *Blood*. 2013;122:1779–1788.
358. Marcella S, Petraroli A, Braile M, et al. Vascular endothelial growth factors and angiopoietins as new players in mastocytosis. *Clin Exp Med*. 2021;21:415–427.
359. Cao TBT, Cha HY, Yang EM, et al. Elevated MRGPRX2 Levels Related to Disease Severity in Patients with Chronic Spontaneous Urticaria. *Allergy Asthma Immunol Res*. 2021;13:498–506.
360. Kirshenbaum AS, Yin Y, Sundstrom JB, et al. Description and Characterization of a Novel Human Mast Cell Line for Scientific Study. *Int J Mol Sci*. 2019;20(22):5520.
361. Akula S, Paivandy A, Fu Z, et al. How Relevant are Bone Marrow-Derived Mast Cells (BMMCs) as Models for Tissue Mast Cells? A Comparative Transcriptome Analysis of BMMCs and Peritoneal Mast Cells. *Cells*. 2020;9(9):2118.
362. Tang XY, Wu S, Wang D, et al. Human organoids in basic research and clinical applications. *Signal Transduct Target Ther*. 2022;7(1):168.
363. Kempuraj D, Selvakumar GP, Thangavel R, et al. Mast Cell Activation in Brain Injury, Stress, and Post-traumatic Stress Disorder and Alzheimer's Disease Pathogenesis. *Front Neurosci*. 2017;11:703.
364. Toms R, Weiner HL, Johnson D. Identification of IgE-positive cells and mast cells in frozen sections of multiple sclerosis brains. *J Neuroimmunol*. 1990;30:169–177.
365. Jennings SV, Finnerty CC, Hobart JS, et al. Mast Cell Diseases in Practice and Research: issues and Perspectives Raised by Patients and Their Recommendations to the Scientific Community and Beyond. *J Allergy Clin Immunol Pract*. 2022;10:2039–2051.